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(57) Abstract

Methods for blocking Ras-induced conditions such as proliferative abnormalities in eukaryote, e.g., mammalian cells. Proteins and mimetics, and their uses, which can block abnormal intracellular signaling often leading to uncontrolled proliferation, e.g., cancers.

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RAS ASSOCIATED GAP PROTEINS

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BACKGROUND OF THE INVENTION

Many proliferative cell abnormalities, e.g., cancers, are caused by alterations in the cellular genome.

10 Mutations can affect the expression or function of genes controlling cell growth and differentiation. See, e.g., Bos (1989) Cancer Research 49:4682-4689. Examples of such oncogenic mutations include members of the Ras family. See, e.g., Mangues et al. (1992) Seminars in Cancer

15 Research 3:229-239. These genes were initially studied as the viral oncogenes of several transforming retroviruses, and their relationship to cellular counterparts was soon recognized. Genes in the Ras family have been shown to be

The Ras family includes three functional genes designated H-ras, K-ras, and N-ras, which encode highly similar proteins. See Barbacid (1987) Ann. Rev. Biochem. 56:779-827. Ras genes from different human tumors were characterized and found to have undergone point mutations leading to constitutive activation, especially codons 12, 13, and 61. These mutant versions are especially potent inducers of tumorigenic or oncogenic transformation. Mutations in the Ras genes may be responsible for as many as 90% of pancreatic adenocarcinomas.

involved in the transduction of extracellular signals and

the control of cellular growth.

The Ras proteins are guanosine triphosphate (GTP) binding proteins, and serve as a molecular switch in signal
transduction controlling the proliferation and
differentiation of cells. The linkage of Ras with the
nucleoside is non-covalent and designated Ras•GXP to
distinguish from a "-" which would indicate a covalent
bond. Two different conformational forms of the protein
exist depending upon the type of guanine nucleoside

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attached to the protein. The Ras•GDP form is an inactive form which does not stimulate the downstream effector, e.g., target protein, to result in functional signal transduction. However, the Ras•GTP conformation is active, e.g., stimulates the effector to transmit an activation signal. Interconversion between the two conformations is enzymatically effected. Conversion from the protein•GDP conformation to the protein•GTP conformation causes activation, and is described as an activation step.

Somatic mutations which constitutively activate Ras, e.g., oncogenic Ras, may contribute to tumorigenesis in up to 30% of human tumors. See, e.g., Bos (1989) Cancer Res. 49:4682- 4689; and Rodenhuis (1992) Seminars in Cancer Biol. 3:241-247. Most anti-cancer drugs currently available are not directed toward specific oncogenes, but rather inhibit even normal cellular processes. These drugs are non-specific and cause severe side-effects, e.g., killing any and all proliferating cell types. Many of these proliferating cells are important for sustaining the organism, e.g., the hematopoietic and immune systems and the intestinal lining. Treatment for proliferative cell conditions, e.g., chemo- or radio-therapy have debilitating side effects due to the nonspecificity of the drugs.

A need exists for means to more directly target
therapeutic reagents to the proper abnormal cells. The
next generations of anti-cancer drugs will be compounds
which specifically target particular oncogenes, e.g., Ras.
Thus, the development of anti-cancer drugs specifically
targeting Ras oncogenes is an important goal to conquer
human malignancies. The present invention provides these
and many other advantages.

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SUMMARY OF THE INVENTION

The present invention provides methods for blocking 5 Ras-induced effects on eukaryotic cells. Different Ras mutations have been demonstrated to cause oncogenic transformation in eukaryotic cells by providing constitutive activation signaling to the cells. Various fragments of GTPase 10 Activating (GAP) proteins have been identified which specifically interact with defined Ras mutants to block signal transduction. These fragments likely function through a mechanism of interacting with the Ras•GTP activated conformation to block the natural interaction of the effector protein. These fragments thus block the 15 constitutive signal transduction which results in Ras induced constitutive effects.

The present invention provides methods of blocking a Ras-induced effect on a cell, comprising a step of introducing a GTPase Activating (GAP) protein to the cell. Ordinarily, the Ras will be an oncogenic Ras or one which substantially lacks GTPase activity. The Ras-induced effect will typically be induction of cell proliferation or transformation. The cell will often be eukaryotic cell, e.g., a mammalian cell, including a human cell. On some embodiments, the step of introducing is by expression of a nucleic acid encoding the GAP protein.

In preferred embodiments, the GAP protein will bind to the Ras protein with a Kd of less than 200 nM. In other embodiments, the GAP protein is selected from: (a) a fragment of a mammalian GAP protein; (b) a fragment of a mammalian NF1-GRD protein; and (c) a homologue or mimetic of (a) or (b). In particular embodiments, the GAP protein is selected from: (a) a fragment of a mammalian GAP protein having a wild type sequence, including a human GAP protein; and (b) a fragment of a mutant mammalian GAP protein having a sequence with an amino acid substitution at a position corresponding to a position 1063 through 1651 of NF1 or the

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corresponding region of other GAP proteins. Many of these substitutions will be a conservative substitution.

In other embodiments, the GAP protein will interact with Ras and block interaction of an effector molecule which binds to Ras at a position corresponding to a position from 32 to 40 or from 59 to 65.

In various preferred embodiments, the GAP protein does not block signal transduction of non-oncogenic Ras. Greater specificity of action can be achieved by identifying the responsible oncogenic Ras and selecting a GAP protein which specifically blocks the identified oncogenic Ras.

The invention also provides methods of treating an oncogenic Ras transformed cell comprising the step of introducing to said cell a GAP protein capable of suppressing the transformation of said cell. Often, the oncogenic Ras transformed cell will be a mammalian cell, including a human cell.

In some embodiments, the GAP protein does not block signal transduction of non-oncogenic Ras. The method can be improved by adding steps of identifying the responsible oncogenic Ras and selecting a GAP protein which blocks transformation by the identified Ras. Preferably, the GAP protein does not block signal transduction of non-oncogenic Ras, e.g., exhibiting specificity.

In addition, the invention provides methods of identifying appropriate GAP proteins useful for treating a mutated Ras-induced condition of a eukaryote cell comprising: (a) identifying the mutated Ras which induces the condition; and (b) screening various GAP variants for specific variants which are capable of blocking the condition. In some embodiments, the eukaryote cell is a mammalian cell, including a human cell. In a preferred embodiment, additional screening is performed to determine which GAP variants have minimal effect on non-mutated Ras effects.

The invention further provides GAP proteins capable of blocking transformation of a cell, where said

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transformation is due to an oncogenic Ras. In some cases, the GAP protein is selected from: (a) a fragment of a mammalian GAP protein; (b) a fragment of a mammalian NF1-GRD protein; and (c) a homologue or mimetic of (a) or (b). In others, the GAP protein is selected from: (a) a fragment of a mammalian GAP protein having a wild type sequence, including a human GAP protein; and (b) a fragment of a mutant mammalian GAP protein having a sequence with an amino acid substitution at a position corresponding to a position from 1063 through 1651 of NF1 or the corresponding region of other GAP proteins. Often the substitution will be a conservative substitution. In other embodiments, the protein interacts with Ras and blocks interaction of an effector molecule which binds to Ras at a position from 32 to 40 or from 59 to 65. Often the cell is a eukaryotic cell, e.g., a mammalian cell, including a human cell.

In preferred embodiments, the oncogenic Ras substantially lacks GTPase activity. In other embodiments, the protein binds to oncogenic Ras with a Kd of less than 200 nM. Mechanistically, the protein may interfere with interaction of Ras•GTP with an effector compound. In another embodiment, the invention provides an isolated nucleic acid encoding a protein normally expressed as a protein as described.

BRIEF DESCRIPTION OF THE DRAWING

Figure 1 shows stimulation of GTPase activity of c-HaRasGly12 and c-Ha-RasVal12 proteins by yeast cell extracts containing wild-type and mutant NF1-GRDs.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

- 10 OUTLINE
 - I. Ras family
 - A. structure and function
 - B. cycling between Ras•GDP and Ras•GTP
- 15 II. GAP proteins; family, mammalian, NF1
 - III. Interaction of Ras and GAP proteins
 - IV. Downstream signal transduction
 - V. Methods
 - A. administering
- 20 B. matching to corresponding Ras
 - C. making compositions, analogues, mimetics
 - I. Ras family
 - A. structure and function
- Ras gene family members are ubiquitous among eukaryotic cells. See, e.g., Barbacid (1987) Ann. Rev. Biochem. 56:779-827. The genes were initially identified and studied as the viral oncogenes of several acute transforming retroviruses. The relationship to human
- cancer was quickly established upon recognition that the retroviral oncogenes were derived from a group of mammalian cellular proto-oncogenes, e.g., endogenous genes which become oncogenic upon mutation.
- Point mutations in the normal endogenous mammalian Ras gene often led to an oncogenic transformed phenotype. Further studies on the locations of the point mutations showed a high frequency at particular hot spots, e.g., codons 12, 13, or 61. Recent technology, e.g., selective hybridization with specific probes, and PCR techniques have
- 40 simplified analysis of specific alterations responsible for

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Ras-induced effects. See Bos (1988) Mutat. Res. 195:255-271.

Extension of interest to the counterparts in non-mammalian systems has shown that these genes play a critical role in transduction of many extracellular signals in cells. Functional and structural data has shown that Ras proteins are GTP-binding proteins involved in transduction of signals in response to extracellular stimuli. The family of Ras proteins can be defined by a combination of functional and structural criteria. See, e.g., Bollag et al. (1991) Ann. Rev. Cell Biol. 7:601-632. Ras-induced effects are the functional consequences of Ras activation.

In mammalian cells, typically the Ras-induced effects will be cell transformation, but may also include differentiation or proliferation effects which fail to satisfy the full criteria for transformation.

The yeast <u>Saccharomyces cerevisiae</u> possesses two members of the Ras family (Rasl and Ras2) which play an important role in cell growth through the regulation of adenylate cyclase. See, e.g., Broach et al. (1990) <u>Adv. Cancer Res.</u> 54:79-139. The Ras-induced effects in yeast show a heat-shock sensitive phenotype.

Members of the Ras family have also been studied in Xenopus laevis; Drosophila melanogaster, Caenorhabditis elegans; and Dictyostelium discoideum. See Bollag et al. (1991) Ann. Rev. Cell Biol. 7:601-632; and Kaziro et al. (1991) Ann. Rev. Biochem. 60:349-400.

Although the Ras-induced effects may be different in

different cells, the relationship in structure often allows
cross species interactions of corresponding proteins in Ras
related pathways. Exploitation of these structural
similarities provide useful means to test interaction of
proteins which normally are never found together with

advantages directed towards ease of testing effects on
various cell sources.

B. cycling between Ras•GDP and Ras•GTP
The Ras proteins have been shown to be GTP-binding
proteins. They can be either in GDP-bound conformation or
a GTP-bound conformation. The GTP-bound conformation is
the active and interacts with an as yet unidentified
effector molecule.

Current models propose that Ras proteins become activated upon stimulation, transduce the signal to an as yet unidentified effector molecule, and subsequently become inactivated. Mutated, e.g., oncogenic, Ras proteins have lost their ability to become inactivated and thus constitutively send a stimulation signal.

Ras is active in its GTP-bound form. The active Ras • GTP complex, which is a non-covalent association, is 15 converted to an inactive Ras-guanosine diphosphate (Ras•GDP) form by an intrinsic GTPase activity found on normal Ras, and which is stimulated by a GTPase Activating (GAP) protein. However, oncogenic Ras lacks the intrinsic GTPase activity and GAP proteins have little, if any, 20 effect on inactivating oncogenic Ras. This substantial lack of GTPase activity in oncogenic Ras will typically be at least 20% less than the normal, more typically at least 35% less, usually at least 50%, more usually at least 60% less, preferably at least 70% less, and more preferably at 25 least 80% or more less than normal Ras.

II. GAP proteins; family, mammalian, NF1 GTPase activities are required to inactivate the Ras•GTP form of the protein in the cycling reaction. A family of proteins stimulating endogenous GTPase activities of Ras proteins have been described which share structural and functional similarities. See Bollag et al. (1991) Ann. Rev. Cell Biol. 7:601-632. Particularly relevant members of the GAP family include yeast and mammalian proteins, including the human neurofibromatosis type 1 (NF1) protein. As used herein, GAP protein refers to a protein which shares structural or functional properties with this family of proteins. Usually, the protein will be a fragment

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shorter than the natural mammalian proteins so far described, normally less than about 600 amino acids, more normally less than about 550 amino acids, ordinarily less than about 500 amino acids, more ordinarily less than about 460 amino acids, usually less than about 420 amino acids, more usually less than about 380 amino acids, typically less than about 350 amino acids, more typically less than about 325 amino acids, preferably less than about 310 amino acids, more preferably less than about 300 amino acids, and in other embodiments, even fewer amino acids, down to 200 or fewer amino acids.

NF1 was first identified as the gene responsible for the pathogenesis of the human genetic disorder, neurofibromatosis type 1. cDNA cloning revealed that the NF1 gene encodes a protein of 2818 amino acids. This putative protein product has a domain showing a significant sequence homology with members of the Ras GTPase-activating protein (GAP) family. See, e.g., Gutmann et al. (1992)

Ann. Neurol. 31:555-561; Xu et al. (1990) Cell 63:835-841; Martin et al. (1990) Cell 63:843-849; and Ballester et al. (1990) Cell 63:851-859. This domain, a fragment of the natural NF1, is often referred herein as NF1 GAP Related Domain (NF1-GRD), and some fragments thereof should have similar activities.

Two yeast <u>Saccharomyces cerevisiae</u> proteins, Iral and Ira2, show particularly high sequence homology to the NF1. Subsequent studies have demonstrated that members of the GAP family, including the GAP-related domain of the NF1 gene product (NF1-GRD; sometimes referred to as NF1 fragment), can stimulate guanosine triphosphatase (GTPase) activity of Ras proteins, i.e., converting Ras•GTP to Ras•GDP, and thereby negatively regulate the activity of Ras.

Two proteins which regulate the activity of Ras

35 proteins are the GTPase activating protein (GAP) and the
protein encoded by NF1, the gene responsible for
neurofibomatosis. type I disease. See Gutmann et al.

(1992) Ann. Neurol. 31:555-561.

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III. Interaction of Ras and GAP proteins

The GAP proteins have been identified as one of the means by which activated Ras proteins are converted into the inactive form. Thus, the physical interaction of the GAP and Ras proteins are important in the understanding of the functional relationship between the entities.

The GAP protein effect on endogenous GTPase activity of RAS has been localized to a fragment of the natural GAP protein, e.g., wild-type sequences. In particular, the catalytic domain has been localized to the carboxy terminal segment of the mammalian GAP proteins. The active portion has been localized to a fragments of less than about 600 amino acids, corresponding to the NF1 amino acids 1063-1651. As such, the functional activities of the GAP proteins would be expected to be localized in this region of the sequence. The sites of GAP interaction with Ras have been proposed to be positions 32-40 and 59-63 of mammalian Ras.

20 The yeast S. cerevisiae possesses two NF1 homologues, Iral and Ira2. The human NF1 is structurally closer to yeast Ira than human GAP and thus would be expected to interact well with the yeast Ras counterpart proteins. This structural similarity is reflected in a functional relationship, as NF1-GRD expressed in yeast cells can 25 complement <u>ira</u>-deficient yeast. In <u>ira</u>-cells, the conversion of Ras•GTP to Ras•GDP is defective, and the cells show a phenotype which is very similar to that of activated Ras mutants, i.e., heat shock-sensitivity. GAP-Related Domain of the NF-1 gene product (NF1-GRD) is a 30 fragment from the NF-1 which can suppress the heatsensitive phenotype of ira , but not of RAS2 Vall9 or RAS2Leu68. This is consistent with the fact that NF1-GRD stimulates GTPase activity of normal but not mutant Ras proteins. Thus, the natural GAP will have blocking effects 35 of Ras functions of normal cells.

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IV. Downstream signal transduction

The biochemical mechanism of signal transduction, or effect, of Ras activation is poorly understood. The structural means by which signal transduction occurs has not been clarified, but it is believed that an effector compound, likely a protein, interacts with Ras•GTP. Genetic analysis of the amino acid positions which affect effector binding have been postulated to include positions 32, 35, 36, 38 and 40. Thus, the effector may well bind near to the same sites of Ras as does the GAP proteins.

This has led to the model that variants of GAP segments may interact with Ras in a fashion which can block effector interaction. This will function to block signal transduction, in a fashion which will inactivate an oncogenically transformed Ras. Moreover, since the different oncogenic Ras forms result from mutations at sites near the GAP and effector interaction sites, variant GAP segments may show great specificity in blocking Rasinduced effects. In particular, the binding affinity of the GAP analogues which block Ras-induced effects are higher than normal GAP binding.

In particular embodiments, the GAP protein, which is intended here to also encompass the concept of protein analogues and mimetics, will preferably be a relatively small polypeptide or analogue, including modified proteins and mimetics. Mimetics include compounds possessing similar molecular shapes sufficient to confer the desired biological property. Various amino acid substitutions may be designed, tested, or screened for activity in blocking Ras-induced functions. These may be effective in blocking effects of many different Ras mutants, or specific Ras variants. The methodology described herein may be useful to define GAP proteins which exhibit high specificity for only interacting with oncogenic, e.g., mutant Ras, and having virtually no effect on natural Ras function. Thus, the GAP proteins provided herein will be highly specific in affecting only oncogenic functions and will be innocuous in cells possessing normal Ras.

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Although the positions of GAP believed to be most important in the interaction with Ras are in the regions of 701-1047 of GAP, the NF1 regions considered most likely to be useful herein will be within the region of 1063-1651 or the corresponding region of other GAP proteins, including 1175-1534, and more specifically in the regions of 1400-1500. Mutations within this region are likely to interact with the Ras in the desired way, particularly in the region of 1421-1461 of NF1 or the corresponding region of other GAP proteins.

Functionally, the useful GAP proteins have high binding affinity for Ras or Ras-like proteins or GAP binding segments thereof. Typically, the GAP protein will exhibit a Kd for Ras, or its oncogenic variant, of less than about 300 nM, more typically less than about 250 nM, usually less than about 200 nM, more usually less than about 150 nM, preferably less than about 100 nM, and more preferably even higher binding affinity. Typically a higher binding affinity will allow effective competitive effect on the effector binding at low concentrations of GAP protein.

IV. Methods

A. administering

25 As described, blocking Ras-induced effects will occur upon proper selection of the GAP protein, e.g., fragments, analogues, and mimetics, and administering such composition to the cell. The GAP protein will be produced, e.g., by recombinant means, as are described in Sambrook et al. 30 (1989) Molecular Cloning: A Laboratory Manual Cold Spring Harbor Press, CSH, N.Y., and Ausubel (1987 and periodic supplements) Current Protocols in Molecular Biology Greene/Wiley, New York; which are each incorporated herein by reference. The GAP protein can be purified and then administered to a patient. These reagents can be combined 35 for therapeutic use with additional active ingredients, e.g., in conventional pharmaceutically acceptable carriers or diluents, along with physiologically innocuous

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stabilizers and excipients. These combinations can be sterile filtered and placed into dosage forms as by lyophilization in dosage vials or storage in stabilized aqueous preparations.

Drug screening using Ras or fragments thereof can be performed to identify compounds having binding affinity. Subsequent biological assays can then be utilized to determine if the compound has intrinsic activity and is therefore a blocker or antagonist in that it blocks the effects of oncogenic Ras. Additional compounds may be screened or designed using the reagents described, or by molecular modeling and structural studies including, e.g., X-ray crystallography, multidimensional NMR, and other techniques. See, e.g., Blundell et al. (1976) Protein Crystallography Academic Press, New York.

The quantities of reagents necessary for effective therapy will depend upon many different factors, including means of administration, target site, physiological state of the patient, and other medicants administered. 20 treatment dosages should be titrated to optimize safety and efficacy. Typically, dosages used in vitro may provide useful guidance in the amounts useful for in situ administration of these reagents. Animal testing of effective doses for treatment of particular disorders will 25 provide further predictive indication of human dosage. Various considerations are described, e.g., in Gilman et al. (eds) (1990) Goodman and Gilman's: The Pharmacological Bases of Therapeutics, 8th Ed., Pergamon Press; and Remington's Pharmaceutical Sciences, 17th ed. (1990), Mack Publishing Co., Easton, Penn.; each of which is hereby 30 incorporated herein by reference. Methods for administration are discussed therein and below, e.g., for oral, intravenous, intraperitoneal, or intramuscular administration, transdermal diffusion, and others. 35 Pharmaceutically acceptable carriers will include water, saline, buffers, and other compounds described, e.g., in the Merck Index, Merck & Co., Rahway, New Jersey. Dosage

ranges would ordinarily be expected to be in amounts lower

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than 100 mM concentrations, typically less than about 10 mM concentrations, usually less than about 100 μ M, preferably less than about 10 μ M, and most preferably less than about 1 μ M, with an appropriate carrier. Slow release

formulations, or slow release apparatus will often be utilized for continuous administration.

The GAP protein may be administered directly to the host to be treated or, depending on the size of the compounds, it may be desirable to conjugate them to carrier proteins such as ovalbumin or serum albumin prior to their administration. Therapeutic formulations may be administered in any conventional dosage formulation. it is possible for the active ingredient to be administered alone, it is preferable to present it as a pharmaceutical formulation. Formulations comprise at least one active ingredient, as defined above, together with one or more acceptable carriers thereof. Each carrier must be both pharmaceutically and physiologically acceptable in the sense of being compatible with the other ingredients and not injurious to the patient. Formulations include those suitable for oral, rectal, nasal, or parenteral (including subcutaneous, intramuscular, intravenous, and intradermal) administration. The formulations may conveniently be presented in unit dosage form and may be prepared by any methods well known in the art of pharmacy. See, e.g., Gilman et al. (eds) (1990) Goodman and Gilman's: The Pharmacological Bases of Therapeutics, 8th Ed., Pergamon Press; and Remington's Pharmaceutical Sciences, 17th ed. (1990), Mack Publishing Co., Easton, Penn.; each of which is hereby incorporated herein by reference. The therapy of this invention may be combined with or used in association with other chemotherapeutic or chemopreventive agents.

Isolation and characterization of these nucleic acids allow use thereof to make variants and mutants. It will also allow production of vector constructs useful, e.g., for gene therapy. See, e.g., Goodnow (1992) "Transgenic Animals" in Roitt (ed.) Encyclopedia of Immunology Academic Press, San Diego, pp. 1502-1504; Travis (1992) Science

256:1392-1394; Kuhn et al. (1991) <u>Science</u> 254:707-710; Capecchi (1989) <u>Science</u> 244:1288; Robertson (1987) (ed.) <u>Teratocarcinomas and Embryonic Stem Cells: A Practical Approach IRL Press, Oxford; and Rosenberg (1992) J. Clinical Oncology</u> 10:180-199; which are each incorporated herein by reference.

B. matching to corresponding Ras

In particular, the present invention allows for simple 10 matching of a therapeutic agent to various oncogenic Ras variants. This can provide highly selective treatment of defined oncogenic conditions with a GAP having highly selected safety and efficacy combinations, virtually tailored to the relatively small number of oncogenic Ras 15 mutations which cause defined proliferative conditions. For example, common variants of oncogenic Ras can be used to screen for GAP fragments which are effective in blocking the oncogenic effects. See, e.g. Kumar et al. (1990) Cancer Res. 52:6877-6884. Either the variants or 20 equivalents thereof can be transformed into a cell, e.g., a yeast cell, and GAP mutants tested for their specific effect on the Ras variants. Once appropriate GAP proteins are identified for each of the common oncogenic Ras mutants, therapeutic reagents can be selected based upon 25 the diagnosed mutant oncogenic Ras responsible for a given abnormality. Diagnosis of the responsible Ras mutation can be performed as described above.

Isolated GAP encoding DNAs can be readily modified by nucleotide substitutions, nucleotide deletions, nucleotide insertions, and inversions of nucleotide stretches. These modifications result in novel DNA sequences which encode these modified GAP proteins, their derivatives, or proteins having the desired anti-oncogenic activity. These modified sequences can be used to produce mutant GAP proteins or to enhance the expression of GAP. Enhanced expression may involve gene

amplification, increased transcription, increased.... translation, and other mechanisms. Such mutant Ras or GAP derivatives include predetermined or site-specific mutations of the respective protein or its fragments. A mutant GAP is a polypeptide otherwise falling within the homology defined by structure and function, but having an amino acid sequence which differs from the corresponding segment of GAP as found in nature, whether by way of an amino acid deletion, substitution, or insertion. Similar proteins and nucleic acids should be 10 available from other warm blooded animals, e.g., mammals and birds. These descriptions are generally meant to encompass species and allelic variants of the GAP proteins, not limited to the specific embodiments 15 discussed.

Although site specific mutation sites are predetermined, mutants need not be site specific. GAP protein or Ras protein mutagenesis can be conducted by making amino acid insertions or deletions.

Substitutions, deletions, insertions, or any combinations may be generated to arrive at a final construct. Insertions include but are not limited to amino- or carboxy- terminal fusions. Random mutagenesis can be conducted at a target codon and the expressed GAP mutants can then be screened for the desired activity. Methods for making substitution mutations at predetermined sites in DNA having a known sequence are well known in the art, e.g., by M13 primer mutagenesis. See also Sambrook et al. (1989) and Ausubel et al. (1987 and Supplements).

The mutations in the DNA normally should not place coding sequences out of reading frames and preferably will not create complementary regions that could hybridize to produce secondary mRNA structure such as loops or hairpins.

The present invention also provides recombinant proteins, e.g., heterologous fusion proteins using segments from these proteins. A heterologous fusion

protein is a fusion of proteins or segments which are naturally not normally fused in the same manner. Thus, the fusion product of an immunoglobulin with a GAP polypeptide is a continuous protein molecule having sequences fused in a typical peptide linkage, e.g., typically made as a single translation product and exhibiting properties derived from each source peptide. A similar concept applies to heterologous nucleic acid sequences.

10 In addition, new constructs may be made from combining similar functional domains from other proteins. For example, Ras-binding or other segments may be "swapped" between different new fusion polypeptides or fragments. See, e.g., Cunningham et al. 15 (1989) Science 243:1330-1336; and O'Dowd et al. (1988) J. Biol. Chem. 263:15985-15992, each of which is incorporated herein by reference. Thus, new chimeric polypeptides exhibiting new combinations of specificities will result from the functional linkage of 20 Ras-binding specificities. For example, the Ras-binding segments from other related proteins may be added or combined with other binding segments from other proteins. The resulting protein will often have hybrid function and properties.

25 The phosphoramidite method described by Beaucage and Caruthers (1981) <u>Tetra</u>. <u>Letts</u>. 22:1859-1862, will produce suitable synthetic DNA fragments. A double stranded fragment will often be obtained either by synthesizing the complementary strand and annealing the strand together under appropriate conditions or by adding the complementary strand using DNA polymerase with an appropriate primer sequence.

The present invention provides means to produce fusion proteins. Various GAP variants may have slightly different functions or biological activities, even though they share significant structural similarities. Dissection of structural elements which effect the various physiological functions or biological activities

provided by the GAP proteins is possible using standard techniques of modern molecular biology, particularly in comparing variants within the related family of GAP proteins. See, e.g., the homolog-scanning mutagenesis technique described in Cunningham et al. (1989) Science 243:1339-1336; and approaches used in O'Dowd et al. (1988) J. Biol. Chem. 263:15985-15992; and Lechleiter et al. (1990) EMBO J. 9:4381-4390; each of which is incorporated herein by reference.

In particular, Ras binding segments can be substituted between proteins to determine what structural features are important in both Ras binding affinity and specificity for the natural or oncogenic Ras. An array of different Ras variants, e.g., allelic, will be used to screen for GAP proteins exhibiting desired properties of interaction with them, e.g., high binding affinity, blocking of effector function by conformational or competitive inhibition, or even forms which can induce GTPase action of the oncogenic Ras.

The specific segments of interaction of GAP with Ras may be identified by mutagenesis or direct biochemical

O The specific segments of interaction of GAP with Ras may be identified by mutagenesis or direct biochemical means, e.g., cross-linking or affinity methods.

Structural analysis by crystallographic or other physical methods will also be applicable.

25 Identification of the similarities and differences between Ras oncogenic variants will lead to new diagnostic and therapeutic reagents or treatments.

Structural studies of the Ras variants will lead to design of new GAP proteins, particularly analogues exhibiting desired effect blocking properties. This can be combined with screening methods to isolate new GAP proteins exhibiting desired spectra of activities. Both the naturally occurring and the recombinant forms of Ras are particularly useful in kits and assay methods which are capable of screening compounds for binding activity to them. Several methods of automating assays have been developed in recent years so as to permit screening of tens of thousands of compounds per year. See, e.g.,

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Fodor et al. (1991) <u>Science</u> 251:767-773, which is incorporated herein by reference and which describes means for testing of binding affinity by a plurality of defined polymers synthesized on a solid substrate. Phage or other libraries of various random polypeptide sequences would also be useful. The development of suitable assays can be greatly facilitated by the availability of large amounts of purified, soluble Ras, either natural or oncogenic, by methods as provided

Expression in other cell types will often result in glycosylation differences in a particular GAP protein. Various mutants may exhibit distinct biological activities based upon structural differences other than amino acid sequence. Differential modifications may be responsible for differential function, and elucidation of the effects are now made possible.

A nucleic acid which encodes a Ras and GAP are readily available, or can be obtained by chemical synthesis, screening cDNA libraries, or by screening genomic libraries prepared from a wide variety of cell lines or tissue samples. See, e.g., Marchuk et al. (1991) Genomics 11:931-940; and nucleic acid and protein data bases, e.g., Protein Identification Resource (PIR), Georgetown University, Washington, D.C., SwissProt and others, see IntelliGenetics, Menlo Park, CA, or the Univ. Wisconsin Biotechnology Center, Madison, Wisconsin.

This DNA can be expressed in a wide variety of host cells for the synthesis of a Ras, GAP. or fragments thereof which can in turn, for example, be used to generate polyclonal or monoclonal antibodies; for construction and expression of modified Ras or GAP molecules; and for structure/function studies. Each GAP can be expressed in host cells that are transformed or transfected with appropriate expression vectors. These molecules can be substantially free of protein or cellular contaminants, other than those derived from the

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recombinant host, and therefore are particularly useful in pharmaceutical compositions when combined with a pharmaceutically acceptable carrier and/or diluent. The GAP, or portions thereof, may be expressed as fusions with other proteins.

Expression vectors are typically self-replicating DNA or RNA constructs containing the desired Ras or GAP gene or its fragments, usually operably linked to suitable genetic control elements that are recognized in a suitable host cell. These control elements are capable of effecting expression within a suitable host. The specific type of control elements necessary to effect expression will depend upon the eventual host cell used. Generally, the genetic control elements can include a prokaryotic promoter system or a eukaryotic promoter expression control system, and typically include a transcriptional promoter, an optional operator to control the onset of transcription, transcription enhancers to elevate the level of mRNA expression, a sequence that encodes a suitable ribosome binding site, and sequences that terminate transcription and translation. Expression vectors also usually contain an origin of replication that allows the vector to replicate independently of the host cell.

The vectors of this invention contain DNA which encodes a useful GAP-like peptide, or a fragment thereof encoding, e.g., an active polypeptide. The DNA can be under the control of a viral promoter and can encode a selection marker. This invention further contemplates use of such expression vectors which are capable of expressing eukaryotic cDNA coding for a GAP in a prokaryotic or eukaryotic host, where the vector is compatible with the host and where the eukaryotic cDNA coding for the GAP is inserted into the vector such that growth of the host containing the vector expresses the cDNA in question. Usually, expression vectors are designed for stable replication in their host cells or for amplification to greatly increase the total number

of copies of the desirable gene per cell. It is not always necessary to require that an expression vector replicate in a host cell, e.g., it is possible to effect transient expression of the GAP in various hosts using vectors that do not contain a replication origin that is recognized by the host cell. It is also possible to use vectors that cause integration of GAP into the host DNA by recombination.

Vectors, as used herein, comprise plasmids, 10 viruses, bacteriophage, integratable DNA fragments, and other vehicles which enable the integration of DNA fragments into the genome of the host. Expression vectors are specialized vectors which contain genetic control elements that effect expression of operably 15 linked genes. Plasmids are the most commonly used form of vector but all other forms of vectors which serve an equivalent function and which are, or become, known in the art are suitable for use herein. See, e.g., Pouwels et al. (1985 and Supplements) Cloning Vectors: A 20 Laboratory Manual, Elsevier, N.Y., and Rodriquez et al. (eds) Vectors: A Survey of Molecular Cloning Vectors and Their Uses, Buttersworth, Boston, 1988, which are incorporated herein by reference.

For purposes of this invention, DNA sequences are operably linked when they are functionally related to each other. For example, DNA for a presequence or secretory leader is operably linked to a polypeptide if it is expressed as a preprotein or participates in directing the polypeptide to the cell membrane or in secretion of the polypeptide. A promoter is operably linked to a coding sequence if it controls the transcription of the polypeptide; a ribosome binding site is operably linked to a coding sequence if it is positioned to permit translation. Usually, operably linked means contiguous and in reading frame, however, certain genetic elements such as repressor genes are not contiguously linked but still bind to operator sequences that in turn control expression.

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Suitable host cells include prokaryotes, lower eukaryotes, and higher eukaryotes. Prokaryotes include both gram negative and gram positive organisms, e.g., E. coli and B. subtilis. Lower eukaryotes include yeasts, e.g., S. cerevisiae and Pichia, and species of the genus Dictyostelium. Higher eukaryotes include established tissue culture cell lines from animal cells, both of non-mammalian origin, e.g., insect cells, and birds, and of mammalian origin, e.g., human, primates, and rodents.

Prokaryotic host-vector systems include a wide variety of vectors for many different species. herein, E. coli and its vectors will be used generically to include equivalent vectors used in other prokaryotes. A representative vector for amplifying DNA is pBR322 or many of its derivatives. Vectors that can be used to express the GAP protein include, but are not limited to, such vectors as those containing the lac promoter (pUC-series); trp promoter (pBR322-trp); Ipp promoter (the pIN-series); lambda-pP or pR promoters (pOTS); or hybrid promoters such as ptac (pDR540). See Brosius et al. (1988) "Expression Vectors Employing Lambda-, trp-, lac-, and Ipp-derived Promoters", in Vectors: A Survey of Molecular Cloning Vectors and Their Uses, (eds. Rodriguez and Denhardt), Buttersworth, Boston, Chapter 10, pp. 205-236, which is incorporated herein by reference.

Lower eukaryotes, e.g., yeasts and Dictyostelium, may be transformed with GAP sequence containing vectors. For purposes of this invention, the most common lower eukaryotic host is the baker's yeast, Saccharomyces cerevisiae. It will be used to generically represent lower eukaryotes although a number of other strains and species are also available. Yeast vectors typically consist of a replication origin (unless of the integrating type), a selection gene, a promoter, DNA encoding the Ras or GAP protein or its fragments, and sequences for translation termination, polyadenylation, and transcription termination. Suitable expression

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vectors for yeast include such constitutive promoters as 3-phosphoglycerate kinase and various other glycolytic enzyme gene promoters or such inducible promoters as the alcohol dehydrogenase 2 promoter or metallothionine promoter. Suitable vectors include derivatives of the following types: self-replicating low copy number (such as the YRp-series), self-replicating high copy number (such as the YEp-series); integrating types (such as the YIp-series), or mini-chromosomes (such as the YCp-series).

Higher eukaryotic cells grown in tissue culture are often the preferred host cells for expression of the GAP protein. In principle, any higher eukaryotic tissue culture cell line is workable, e.g., insect baculovirus 15 expression systems, whether from an invertebrate or vertebrate source. However, mammalian cells are often preferred. Transformation or transfection and propagation of such cells has become a routine procedure. Examples of useful cell lines include HeLa 20 cells, Chinese hamster ovary (CHO) cell lines, baby rat kidney (BRK) cell lines, insect cell lines, bird cell lines, and monkey (COS) cell lines. Expression vectors for such cell lines usually include an origin of replication, a promoter, a translation initiation site, 25 RNA splice sites (if genomic DNA is used), a polyadenylation site, and a transcription termination site. These vectors also usually contain a selection gene or amplification gene. Suitable expression vectors may be plasmids, viruses, or retroviruses carrying 30 promoters derived, e.g., from such sources as from adenovirus, SV40, parvoviruses, vaccinia virus, or cytomegalovirus. Representative examples of suitable expression vectors include pCDNA1 (Invitrogen, San Diego, CA); pCD, see Okayama et al. (1985) Mol. Cell 35 Biol. 5:1136-1142; pMClneo Poly-A, see Thomas et al. (1987) Cell 51:503-512; and a baculovirus vector such as

pAC 373 or pAC 610.

It may be desired to express a GAP polypeptide in a system which provides a specific or defined glycosylation pattern. In this case, the usual pattern will be that provided naturally by the expression 5 system. However, the pattern will be modifiable by exposing the polypeptide, e.g., an unglycosylated form, to appropriate glycosylating proteins introduced into a heterologous expression system. For example, the GAP gene may be co-transformed with one or more genes encoding mammalian or other glycosylating enzymes. Using this approach, certain mammalian glycosylation patterns will be achievable in prokaryote or other cells.

15 The broad scope of this invention is best understood with reference to the following examples, which are not intended to limit the invention in any manner.

EXAMPLES

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In these studies, a yeast Ras system was used to isolate NF1-GRD mutants which can suppress specifically the activity of oncogenic Ras. Yeast cells carrying activated mutations in Ras (such as RAS2 Vall9 and RAS2 Leu68) are defective in responding to environmental conditions, and show a variety of phenotypes including a heat shocksensitive phenotype.

First, a pool of randomly mutagenized NF1-GRD genes were screened to obtain suppressors of a specific yeast oncogenic-type Ras, RAS2 Val19. Next, these mutant NF1-GRDs were shown to be capable of inhibiting v-Ras-induced transformation in mammalian cells. These results demonstrated that this unique yeast method provides a powerful screening system to obtain anti-Ras NF1-GRD The mutants of NF1-GRD most likely bind tightly with the oncogenic, e.g., mutated, Ras proteins to sequester the latter proteins from the signal transduction for normal cell growth. Detailed analysis of the

structures involved in the interaction between mutant NF1-GRDs and Ras will enable testing of compounds, e.g., analogues and mimetics, which can mimic the action of NF1-GRDs, and inhibit specifically transforming Ras activity.

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EXAMPLE 1: Preparation of pKP11

A plasmid pKP11, which expresses a domain of NF1
(amino acid residues 1063-1651; the numbers of amino acid
residues are referred to according to Marchuk et al. (1991)
Genomics 11:931-940, and a yeast strain carrying RAS2Val19
mutation were used to obtain mutant NF1-GAP Related Domains
(GRDs) which can suppress the phenotype of activated Ras.
In a previous study, this plasmid was shown to suppress
15 ira2 but not RAS2Val19. The plasmid was randomly
mutagenized by treatment with hydroxylamine in vitro, and a
pool of mutagenized DNAs was transformed into RAS2Val19
cells. Subsequently, about 2 x 105 independent colonies
were screened for heat shock resistance.

20 Wild-type NF1-GRD was cloned into the yeast expression vector pKT10 which contains glyceraldehyde-3-phosphate dehydrogenese promoter, a replication origin derived from 2 μm, and <u>URA3</u> as a selection marker to yield pKP11. One hundred micrograms of pKP11 DNA was mutagenized by 25 hydroxylamine in vitro as described previously (Rose et al. (1987) Cell 48:1047-1060), and transformed into a S. cerevisiae strain, TK161-R2V-D which carries RAS2Val19 mutation. See Tanaka et al. (1989) Mol. Cell. Biol. 9:757-768; and Tanaka et al. (1990) Mol. Cell. Biol. 10:4303-4313. About 2 x 10^5 colonies were grown on selection 30 plates, and the plates were heated at 57 °C for 15 minutes. The resultant plates were incubated at 30 °C for 4 days, and growing colonies were selected. The heat shocksensitivity of these colonies were checked, and 12 clones 35 were selected at this stage. Plasmid DNAs were recovered from these cells, re-transformed into TK161-R2V-D, and phenotypic reversion was examined.

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Twelve positive colonies were obtained in the initial screening. Subsequently, two clones, NF201 (SEQ ID NO: 1) and NF204 (SEQ ID NO: 2), which had a relatively strong suppression activity for $RAS2^{Vall9}$, were selected, and subjected to further analysis.

EXAMPLE 2: Effect of Mutant NF1-GRDs on yeast cells

The effects of NF201 (SEQ ID NO: 1) and NF204 (SEQ ID NO: 2) were tested on different alleles of activated RAS2Val19 in yeast cells (Table 1). Wild-type NF1-GRD could weakly revert the phenotype of RAS2Leu68, but was totally ineffective on RAS2Val19 and RAS2Ser41. Mutant NF201 suppressed the heat shock-sensitive phenotype of all three alleles of RAS2 examined, including RAS2Val19, RAS2Leu68, and RAS2Ser41 (Tanaka et al. (1992) Mol. Cell. Biol. 21:631-637). On the other hand, NF204 preferentially suppressed RAS2Val19 but not the other two alleles. These results indicate that NF201 and NF204 possess distinct properties as suppressors of activated Ras in a Ras-

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specific manner.

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Suppression of the heat-sensitive phenotypes of various activated alleles of RAS2 by mutant NF1-GRD. A wild-type S. cerevisiae strain, RAY-3A-D, harboring a combination of RAS2 plasmids (YCp-RAS2 Vall9, -RAS2 Leu68, and -RAS2Ser41; Tanaka et al. (1992) Mol. Cell. Biol. 12:631-637) and NF1-GRD plasmids, was subjected to heat shock assay. The ability of each NF1-GRD plasmid to suppress the heat-sensitive phenotype was scored: +++, strong suppression; ++, intermediate suppression; +, weak suppression; -, no detectable suppression. The 10 complementation activity in ira2 cells (KT63-2B-D; Tanaka et al. (1989) Mol. Cell. Biol. 9:757-768; Tanaka et al. (1990) Mol. Cell. Biol. 10:4303-4313), which reflects the activity of these NF1-GRDs on wild-type RAS2 (RAS2Wt), was 15 also scored, and is shown in the table.

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RAS2 allele

NF1-GRD	RAS2Val19	RAS2Leu68	RAS2Ser41	RAS2Wt
NF201	+++	+++	++	+++
NF204	+++	+	-	+++
NF1 (wild-type)	-	+ ,	-	+++
	NF201 NF204	NF201 +++ NF204 +++	NF201 +++ +++ NF204 +++ +	NF201 +++ +++ ++ NF204 +++ + -

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Interestingly, these two mutant NF1-GRDs could suppress <u>ira2</u> cells, in which normal Ras proteins are activated, to the same extent as wild-type NF1-GRD, suggesting that NF201 and NF204 retain the ability to stimulate GTPase activity of normal Ras.

The entire region of mutant NF1-GRDs were sequenced to identify mutations in NF201 and NF204, and the sequences compared the sequences with that of wild-type NF1-GRD. In both NF201 and NF204, single nucleotide changes were found in the DNA sequences. In NF201 (SEQ ID NO: 1), the codon TTC for Phe at residue 1434 was changed to TTA coding for

Leu, while in NF204 (SEQ ID NO: 2), the codon AAG for Lys at residue 1436 was replaced by AGA coding for Arg.

Although both mutation sites are located in one of the most conserved regions of the GAP-related domain (see Xu et al. (1990) <u>Cell</u> 63:835-842; Martin et al.(1990) <u>Cell</u> 5 63:843-850; and Ballester et al. (1990) Cell 63:851-859), the amino acid residues at these sites (Phe at position 1434, and Lys at position 1436) are not strictly conserved among the members of the GAP family (Figure 1). residue at 1434 in NF1 is conserved in yeast Ira2 (SEQ ID 10 NO: 4) protein, but it is replaced by other residues in Iral (SEQ ID NO: 3), GAP (SEQ ID NO: 5), and Gapl (SEQ ID NO: 6). On the other hand, Lys residue at 1346 is conserved among NF1, Iral, GAP, and Gap1, but Ira2 contains Arg at the corresponding site. Recently, two independent 15 studies have demonstrated that Lys at position 1423 in NF1-GRD, which is located just 11 and 13 amino acids upstream of the mutation sites of NF201 and NF204, respectively, is important for the structure and function of NF1. the substitution of Glu for Lys at position 1423 has been 20 identified in some human tumors as well as in a family of neurofibromatosis patients (Li et al. (1992) Cell 69:275-The GAP activity of this mutant NF1-GRD was 200- to 400-fold lower than that of the wild-type NF1-GRD. also reported that the substitution of Met for Lys at the 25 same position resulted in a decrease in stability of the protein (Wiesmuller et al. (1992) J. Biol. Chem. 267:10207-10219). Thus, the amino acid residues at 1423, 1434 and 1436, and their surrounding sequence, are likely to be 30 important for the structure and/or function of NF1 proteins.

EXAMPLE 3: Effect of mutant NF1-GRDs in mammalian cells

The effect of these mutant NF1-GRDs in mammalian cells
35 was investigated. The cDNA fragments of the wild-type and
mutant NF1-GRDs were recloned into a mammalian expression
vector, and transfected into cell lines.

The size of the NF1-GRD protein transiently expressed in Cos7 cells was checked. Western blot analysis with an anti-NF1-GRD anti-serum (see Hattori et al. (1992) Oncogene 7:481-485) identified a protein band of an apparent molecular mass of 67-68 kDa in the cells transfected with NF1-GRD plasmids but not with the control vector. This suggests that the protein of about 67 kDa was translated starting from the internal Met residue at position 1073 of NF1 cDNA.

The anti-Ras activities of mutant NF1-GRDs were 10 examined for their effects on v-Ras-induced transformation. The above plasmids expressing NF1-GRD were cotransfected with pSV2neo into DT cells, a v-Ki-ras-transformed NIH3T3 derivative, and the ability to induce morphological reversion of the cells was examined. As shown in Table 2, 15 transfection of the plasmids expressing NF201 and NF204 could induce flat reversion at dramatically high frequencies (8-9% of total G418-resistant colonies). The frequency was even higher than that obtained by 20 transfection of a Krey-1 plasmid which has been shown to possess anti-oncogenic activity in DT cells (Kitamura et al. (1990) Proc. Natl Acad. Sci. USA 87:4284-4288). Under the same conditions, the wild-type NF1-GRD could also induce flat reversion of DT cells, although it was 5 to 6 times less potent than mutant clones. This is particularly 25 interesting since a previous study has shown that overexpression of GAP inhibited normal c-Ha-Ras- but not v-Ha-Ras-induced transformation (see Zhang et al. (1990) Nature 346:754-756).

No revertant of DT cells could be obtained from transfectants of the GAP plasmid (Table 2). This difference may be due to the fact that NF1-GRD possesses a much higher affinity for Ras proteins than GAP. These results clearly demonstrate that mutant NF1-GRDs possess transformation-suppressor activity against oncogenic Ras.

Table 2. Induction of morphological reversion of v-Rastransformed cells by mutant NF1-GRD. DT cells were cotransfected with 20 μg of NF1-GRD plasmids and 2 μg of pSV2neo as described by Kitamura et al. (1990) Proc. Natl Acad. Sci. USA 87:4284-4288, and transfectants were selected in a medium containing 0.5 mg/ml G418. Since pKrey-1 plasmid itself contained the neo gene, 2 μg of the plasmid was cotransfected with 20 μg of pEF-BOS (the vector for NF1-GRD). The pEF-GAP contained rat full-length GAP cDNA in pEF-BOS. Frequency of reversion in DT cells is defined as the ratio (%) of morphologically flat cell colonies to total G418-resistant colonies. N.D.: not determined.

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20 Flat colonies/G418-resistant colonies

25	transfected DNA	d Exp.1	Exp.2	Exp.3	ratio	
						
	pEF-BOS	0/1155 (<0.1)	2/1279 (0.1)	3/878 (0.4)	0.1	
30	per-nf1	20/1522 (1.3)	26/1151 (2.3)	15/1004 (1.5)	1.7	
	per-NF201	86/1190 (7.2)	61/691 (8.8)	34/356 (9.6)	8.0	
35	pEF-NF204	40/448 (8.9)	46/426 (10.8)	24/350 (6.9)	9.0	
	pEF-GAP	N.D.	0/856 (<0.1)	0/561 (<0.2)	<0.1	
	pK <u>rev</u> -1	N.D.	26/1385 (1.9)	15/736 (2.0)	1.9	
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EXAMPLE 4: Biochemical properties of the mutant NF1-GRDs

The biological properties of the mutant NF1-GRDs were 5 studied to understand the molecular mechanism of antioncogenic activity. Extracts were prepared from yeast cells expressing wild-type and mutant NF1-GRDs, and GTPasestimulating activity was measured in vitro by using recombinant c-Ha-Ras proteins as substrates. Recombinant c-Ha-RasGly12 (A) or c-Ha-RasVall2 (B) proteins were loaded 10 with $[\gamma-32P]GTP$ (30 Ci/mmol) in buffer B (50 mM tris-HCl [pH 7.4], 50 mM KCl, 1 mM MgCl₂, 2.5 mM EDTA, and 0.2 mg/ml BSA) at 30 'C for 10 minutes. The reaction was stopped by the addition of MgCl₂ to the final concentration of 7 mM. 15 Yeast cell extracts were prepared from wild-type yeast cells, RAY-3A-D, carrying various NF1-GRD plasmids. Cells grown to the stationary phase were collected, and disrupted with acid-washed glass beads (0.5 mm diameter) in buffer A (50 mM tris-HCl [pH 7.4], 100 mM KCl, 5 mM MgCl₂, 2 mM DTT, 20 2 mM PMSF, 1 mM benzamidine, and 10 µg/ml of each of pepstatin A, aprotinin, and leupeptin. The crude extract was clarified twice by centrifugation at 2000 x g for 20 The resultant supernatants were then mixed with an aliquot of Ras•[γ -32P]GTP mixture, and incubated at 30 25 At the indicated time point, an aliquot was filtered through a nitrocellulose membrane, and radioactivity retained on the membrane was counted. The final concentrations of yeast extract proteins and Ras $(\gamma^{-32}P)$ GTP were 1 mg/ml and 11.5 nM, respectively. The cell extracts 30 assayed were from the cell carrying the following plasmids: ., wild-type NF1-GRD; o, NF201; Δ, NF204; [solid square], vector alone; or [open square], buffer A plus 1 mg/ml BSA. Two mutant NF1-GRDs, NF201 and NF204, stimulated the GTPase activity of c-Ha-Ras^{Gly12} to the same extent as wild-type 35 NF1-GRD (Figure 1).

This is consistent with the observation that NF201 and NF204 can effectively complement <u>ira2</u> in yeast (see Table

1). On the other hand, the same extracts were not able to stimulate the GTPase activity of $c\text{-Ha-Ras}Vall2}$ under these experimental conditions. This suggests that the anti-oncogenic activity of the mutant NF1-GRD is not due to the stimulation of the slow GTPase of oncogenic Ras proteins.

The members of the GAP family negatively regulate the activity of Ras by stimulating intrinsic GTPase activity of normal Ras proteins. Thus, NF1 can potentially act as a specific block of effector function by normal Ras. 10 However, oncogenic Ras lacks the intrinsic GTPase activity, and thus, natural GAP sequences cannot stimulate the inactivation of the activated oncogenic Ras. NF1-GRD suppresses the heat shock-sensitive phenotype of ira cells, but not the same phenotype of activated mutants 15 of Ras, e.g., RAS2 Val19 and RAS2 Leu68 which correspond to mammalian oncogenic Ras, ras Vall2 and ras Leu61. respectively. Various mammalian oncogenic Ras mutants may be simulated by corresponding mutations in yeast Ras 20 proteins. These observations lead to a model which is useful for testing interaction of Ras variants with GAP variants, and which predicts useful blocking or reversal of mutant or oncogenic Ras-induced effects.

25 A model of anti-oncogenic activity of mutant NF1-GRD consistent with these observations is that the mutant NF1-GRD has higher affinity for oncogenic Ras.GTP as compared to the wild-type NF1-GRD. As discussed above, the GAP binding region, and the effector binding regions on the Ras 30 protein are in close physical proximity. As such, mutant NF1-GRD binding to oncogenic Ras, e.g., high affinity binding, could form an irreversible NF1.Ras.GTP complex. This could prevent interaction with putative downstream effector molecules, e.g., by conformational changes or 35 competition. The oncogenic Ras would be sequestered from signal transduction pathways. Two observations support this hypothesis. First, as shown in Table 1, weak but significant phenotypic reversion of RAS2 Leu68 by wild-type

NF1-GRD was observed. A previous study (Bollag et al. (1991) Nature 351:576-579) showed that the mammalian RasLeu61 protein (corresponding to yeast RAS2Leu68) has a much higher affinity for NF1-GRD than the wild-type or Vall2-form of Ras. The high affinity binding between 5 RAS2Leu68 and wild-type NF1-GRD can explain the phenotypic suppression. Likewise, this model can also explain the differences in transformation-suppressor activities among GAP, wild-type NF1-GRD, and mutant NF1-GRDs. Table 2 shows that wild-type NF1-GRD, but not GAP, can suppress 10 transformation by v-Ras; two mutant NF1-GRDs are more potent suppressors than wild-type NF1-GRD. This order of potency as transformation suppressors may reflect the relative affinity for Ras proteins; that is, wild-type NF1-15 GRD has 20 times higher affinity for Ras than GAP (see Martin et al. Cell 63:843-850); mutant NF1-GRDs may have even greater affinities. In relation to this, it should be noted that Ballester et al. (1990) Cell 63:851-859 previously observed the inhibitory effect of wild-type NF1-GRD but not of GAP on c-Ha-Ras Vall2 expressed in yeast 20 cells. This is consistent with the observation that wildtype NF1-GRD can weakly suppress v-Ras-transformation in mammalian cells. The second observation supporting this model is that NF201 can suppress the activity of not only RAS2 Val19 and RAS2 Leu68, but also RAS2 Ser41. It has been 25 shown that Ser41 mutation (corresponding to Ser34 of human Ras), which is located in the so-called "effector region," disrupts the effective binding of Ras2 proteins to yeast Ira proteins as well as NF1-GRD and GAP (Tanaka et al. 30 (1992) Mol. Cell. Biol. 12:631-637). Thus, the fact that NF201 can inhibit the activity of RAS2 Ser41 strongly suggests that the mutation in NF201 restores the interaction between RAS2Ser41 and NF1-GRD. Comparison of the relative affinities of wild-type and mutant NF1-GRDs 35 for oncogenic Ras proteins should provide a test for this model. This model predicts that highly specific reagents could be produced having specificity only for blocking

oncogenic Ras effects while having virtually no effects on normal Ras.

In summary, the data presented herein demonstrated that NF1-GRDs with single amino acid substitutions can suppress the biological activity of oncogenic Ras. According to the proposed model, mutant NF1-GRDs could inhibit specifically oncogenic but not normal Ras. case of normal Ras • GTP, bound GTP would be rapidly hydrolyzed to GDP upon interaction with NF1-GRD, and NF1-10 GRD would be released from Ras.GDP. In this study, a mutant NF1-GRD was expressed as a protein of 578 amino acids, which is still a substantially large protein. yeast screening system described will allow determination of the minimum fragment of NF1-GRD which retains anti-15 oncogenic activity. This approach will allow development of Ras-specific anti-oncogenic compounds.

All references cited herein are incorporated herein by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.

Many modifications and variations of this invention can be made without departing from its spirit and scope, as will be apparent to those skilled in the art. The specific embodiments described herein are offered by way of example only, and the invention is to be limited only by the terms of the appended claims, along with the full scope of equivalents to which such claims are entitled.

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SEQUENCE LISTING

5		
	(1) GENE	RAL INFORMATION:
10	(i)	APPLICANT: Schering Corp.
10	(ii)	TITLE OF INVENTION: RAS Associated GAP Protein
	(iii)	NUMBER OF SEQUENCES: 2
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20		(D) STATE: New Jersey (E) COUNTRY: USA
		(F) ZIP: 94304-1104
25	(v)	COMPUTER READABLE FORM: (A) MEDIUM TYPE: Floppy disk (B) COMPUTER: Macintosh (C) OPERATING SYSTEM: 6.0.8 (D) SOFTWARE: Microsoft Word 5.1a
30	(vii	PRIOR APPLICATION DATA: (A) APPLICATION NUMBER: US 08/004,824 (B) FILING DATE: 15-JAN-1993 (C) CLASSIFICATION:
35	(viii)	ATTORNEY/AGENT INFORMATION: (A) NAME: Lunn, Paul G. (B) REGISTRATION NUMBER: 32,743 (C) REFERENCE/DOCKET NUMBER: DX0352 PCT
40	(ix)	TELECOMMUNICATION INFORMATION: (A) TELEPHONE: (201)822-7255 (B) TELEFAX: (201)822-7039
45	(2) INFO	RMATION FOR SEQ ID NO:1:
45		SEQUENCE CHARACTERISTICS: (A) LENGTH: 2485 amino acids (B) TYPE: amino acid
50		(C) STRANDEDNESS: single (D) TOPOLOGY: linear
	(ii)	MOLECULE TYPE: protein
55	(vi)	ORIGINAL SOURCE: (A) ORGANISM: Homo sapiens
60	(ix)	FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 5649380

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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30	Met 465	Glu	Asp	Gly	Gln	Ala 470	Ala	Glu	Ser	Leu	His 475	Lys	Thr	Ile	Val	Lys 480
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	His	Ala	Ile 675	Gln	Ile	Lys	Thr	Lys 680	Leu	Cys	Gln	Leu	Val 685	Glu	Val	Met
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	Ser 945	Leu	Ala	Ser	Lys	11e 950	Met	Thr	Phe	Cys	Phe 955	Lys	Val	Tyr	Gly	Ala 960
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25	Ser	Ser	Asp	Trp 980	Gln	His	Val	Ser	Phe 985	Glu	Val	Asp	Pro	Thr 990	Arg	Leu
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5	Asp Thr His Trp Ser Ser Leu Asn Leu Thr Ser Ser Lys Phe Glu Glu 1205 1210 1215	l
	Phe Met Thr Arg His His Gln Val His Glu Lys Glu Glu Phe Lys Ala 1220 1225 1230	
10	Leu Lys Thr Leu Ser Ile Phe Tyr Gln Ala Gly Thr Ser Lys Ala Gly 1235 1240 1245	
15	Asn Pro Ile Phe Tyr Tyr Val Ala Arg Arg Phe Lys Thr Gly Gln Ile 1250 1255 1260	
	Asn Gly Asp Leu Leu Ile Tyr His Val Leu Leu Thr Leu Lys Pro Tyr 1265 1270 1275 128	
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	Gly Leu Lys Gly Ser Lys Arg Leu Val Phe Ile Asp Cys Pro Gly Lys 1345 1350 1355 1360)
35	Leu Ala Glu His Ile Glu His Glu Gln Gln Lys Leu Pro Ala Ala Thr 1365 1370 1375	
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	Leu Asn Asp Ile Tyr Tyr Ala Ser Glu Ile Glu Glu Ile Cys Leu Val 1425 1430 1435 1440)
50	Asp Glu Asn Gln Phe Thr Leu Thr Ile Ala Asn Gln Gly Thr Pro Leu 1445 1450 1455	
	Thr Phe Met His Gln Glu Cys Glu Ala Ile Val Gln Ser Ile Ile His 1460 1465 1470	•
55	Ile Arg Thr Arg Trp Glu Leu Ser Gln Pro Asp Ser Ile Pro Gln His 1475 1480 1485	
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5	Arg Ser Ser Tyr Arg Asp Arg Ser Phe Ser Pro Gly Ser Tyr Glu Arg 1825 1830 1835 1840 Glu Thr Phe Ala Leu Thr Carl
	Glu Thr Phe Ala Leu Thr Ser Leu Glu Thr Val Thr Glu Ala Leu Leu 1845 1850 1855
10	Glu Ile Met Glu Ala Cys Met Arg Asp Ile Pro Thr Cys Lys Trp Leu 1860 1865 1870
	Asp Gln Trp Thr Glu Leu Ala Gln Arg Phe Ala Phe Gln Tyr Asn Pro 1875 1880 1885
15	Ser Leu Gln Pro Arg Ala Leu Val Val Phe Gly Cys Ile Ser Lys Arg 1890 1895 1900
•	Val Ser His Gly Gln Ile Lys Gln Ile Ile Arg Ile Leu Ser Lys Ala 1905 1910 1915 1920
20	Leu Glu Ser Cys Leu Lys Gly Pro Asp Thr Tyr Asn Ser Gln Val Leu 1925 1930 1935
25	Ile Glu Ala Thr Val Ile Ala Leu Thr Lys Leu Gln Pro Leu Leu Asn 1940 1945 1950
	Lys Asp Ser Pro Leu His Lys Ala Leu Phe Trp Val Ala Val Ala Val 1955 1960 1965
30	Leu Gln Leu Asp Glu Val Asn Leu Tyr Ser Ala Gly Thr Ala Leu Leu 1970 1975 1980
	Glu Gln Asn Leu His Thr Leu Asp Ser Leu Arg Ile Phe Asn Asp Lys 1985 1990 1995 2000
35	Ser Pro Glu Glu Val Phe Met Ala Ile Arg Asn Pro Leu Glu Trp His 2005 2010 2015
40	Cys Lys Gln Met Asp His Phe Val Gly Leu Asn Phe Asn Ser Asn Phe 2020 2025 2030
	Asn Phe Ala Leu Val Gly His Leu Leu Lys Gly Tyr Arg His Pro Ser 2035 2040 2045
45	Pro Ala Ile Val Ala Arg Thr Val Arg Ile Leu His Thr Leu Leu Thr 2050 2055 2060
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	Thr			2260)				2265	5				2270)	
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	Asn 2305					2310)				2315	i				2320
35	Pro				2325					2330					2335	5
40.	His			2340		•			2345	;				2350)	
40.	Gly		2355					2360)	_			2365	;		
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	Ile . 2385		Thr	Tyr		Pro 2390		Ile	Asp		Glu 2395		Ser	Glu	Glu	Ser 2400
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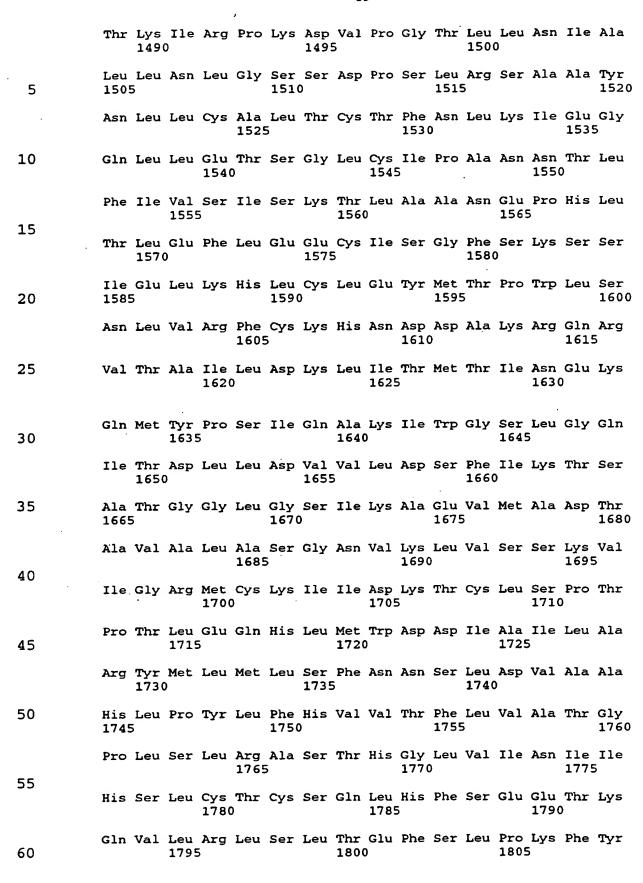
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10	(i)	(A (B (C) LE) TY) ST	E CH NGTH PE: (RAND)	: 24 amin EDNE	85 au o ac SS:	mino id sing	aci	ds							
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20	(vi)			L SOI GANI:			sap	iens								
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25	Ala	Gly	Cys	Ser 340	Gly	Thr	Pro	Pro	Ile 345	Cys	Arg	Gln	Ala	Gln 350	Thr	Lys
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	Lys	Trp 450	Glu	Gln	Ala	Thr	Lys 455	Leu	Ile	Leu	Asn	Tyr 460	Pro	Lys	Ala	Lys
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٠.		Lys	Phe 610	Phe	Asp	Ser	Gln	Gly 615	Gln	Val	Leu	Leu	Thr 620	Asp	Thr	Asn	Thr
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		Phe	Leu 770	Lys	Tyr	Phe	Thr	Leu 775	Phe	Met	Asn	Leu	Leu 780	Asn	Asp	Cys	Ser
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ر	_	Asn	Leu	Leu	Asn 820	Ala	Asn	Val	Asp	Ser 825	Gly	Leu	Met	His	Ser 830	Ile	Gly
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55	Arg	Arg	Phe		Leu 1125		lle	Ala	Ser	Asp 1130		Pro	Thr	Ser	Asp 1135	
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	1250)		1255	Arg Arg	126	0	
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2 5		1300)		Leu Ser 1305		1310)
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4 5	Leu Ala Ala His Gln Val	Leu Glu 1380 Lys Asp 1395 Thr Ser	Glu Asp Thr Lys Ala Glu	His Glu Leu Lys . Val Ser 1400 Arg Thr	1370 Val Phe 1385 Ile Lys Lys Val	Lys Leu His Asn Val Gly Leu Gly 1420	Ala Leu 1390 Ser Thr 1405 Gln Ser	Ala Thr 1375 Lys Leu Ala Val Val Phe
	Leu Ala Ala His Gln Val 1410 Leu Asn 1425	Leu Glu 1380 Lys Asp 1395 Thr Ser	Glu Asp Thr Lys Ala Glu Tyr Tyr 1430	His Glu Leu Lys Val Ser 1400 Arg Thr 1415 Ala Ser	Val Phe 1385 Ile Lys Lys Val	Lys Leu His Asn Val Gly Leu Gly 1420 Glu Glu 1435	Ala Leu 1390 Ser Thr 1405 Gln Ser	Ala Thr 1375 Lys Leu Ala Val Val Phe Leu Val 1440
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4 5	Leu Ala Ala His Gln Val 1410 Leu Asn 1425 Asp Glu Thr Phe	Leu Glu 1380 Lys Asp 1395 Thr Ser Asp Ile Asn Gln Met His 1460	Glu Asp Thr Lys Ala Glu Tyr Tyr 1430 Phe Thr 1445 Gln Glu	His Glu Leu Lys Val Ser 1400 Arg Thr 1415 Ala Ser Leu Thr Cys Glu	Val Phe 1385 Ile Lys Lys Val Glu Ile	Lys Leu His Asn Val Gly Leu Gly 1420 Glu Glu 1435 Asn Gln Val Gln	Ala Leu 1390 Ser Thr 1405 Gln Ser Ile Cys Gly Thr Ser Ile 1470	Ala Thr 1375 Lys Leu Ala Val Val Phe Leu Val 1440 Pro Leu 1455 Ile His



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		Gln 1970	0				1975	5				1980)			
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40		Lys			2009	5		•		2010)				2015	5
		Phe		2020)				2025	5				2030)	
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		2050 Val)				2055	5				2060)			
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55		Arg			2085	5				2090					2095	5
	Ser	Met				Pro	Met	Asp	2105 Thr		Pro	Ile	His	2110 His		Asp
60	Pro	Ser	2115 Tyr		Thr	Leu				Gln					Pro	Lys
		2130)				2135	5				2140	١			

	Gly Ser Glu Gly Tyr Leu Ala Ala Thr Tyr Pro Thr Val Gly Gln 12145 2150 2155 2	Thr 2160
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	Leu Ile Ser Asp Thr Lys Ala Pro Lys Arg Gln Glu Met Glu Ser G 2195 2200 2205	Sly
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	Met Glu Thr Gln Arg Ile Ser Ser Ser Gln Gln His Pro His Leu A 2225 2230 2235 2	rg 240
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	Val Lys Tyr Thr Thr Asp Glu Phe Asp Gln Arg Ile Leu Tyr Glu Tyr 2275 2280 2285	yr
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35	Asn Leu Leu Asp Ser Lys Ile Asn Thr Leu Leu Ser Leu Cys Gln As 2305 2310 2315 23	sp 320
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	Leu Leu Thr Pro Thr Ser Pro Tyr Pro Pro Ala Leu Gln Ser Gln Le 2405 2410 2415	eu
55	Ser Ile Thr Ala Asn Leu Asn Leu Ser Asn Ser Met Thr Ser Leu Al 2420 2425 2430	la
	Thr Ser Gln His Ser Pro Ala Ser Leu Pro Cys Ser Asn Ser Ala Va 2435 2440 2445	1
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5

52

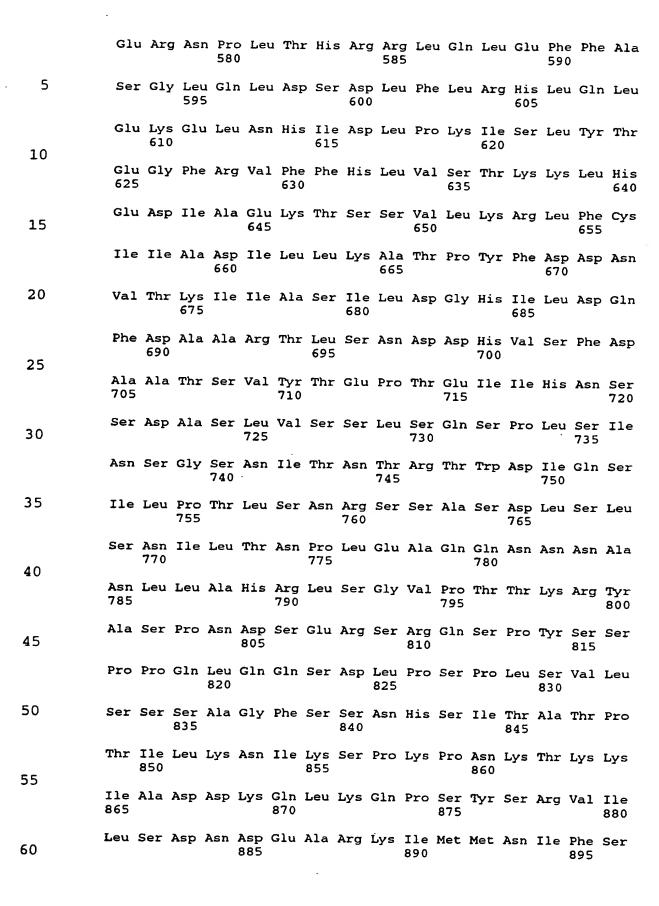
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Ser Ala Ser Gln Val 2485

(2) INFORMATION FOR SEQ ID NO:3:

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5	·(i)	(A (B (C) LE) TY) ST	NGTH PE: RAND	: 29 amin EDNE	TERI 38 a o ac SS: line	mino id sing	aci	ds							
10	(ii)	MOL	ECUL	E TY	PE:	prot	ein									
	(vi)					: Sacc	haro	myce	s ce	revi	siae					
15	(xi)	SEQ	UENC:	E DE	SCRI	PTIO	N: S	EQ I	D NO	:3:						
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	Lys	Val	Leu	Gln 20	Asn	Met	Ser	His	His 25	Leu	Ser	Gly	Ser	Ala 30	Thr	Ile
25	Ser	Lys	Ser 35	Ser	Ile	Leu	Pro	Asp 40	Ser	Gln	Glu	Phe	Leu 45	Gln	Lys	Arg
	Asn	Tyr 50	Pro	Ala	Tyr	Thr	Gl u 5 5	Lys	Ile	Asp	Leu	Thr 60	Ile	Asp	Tyr	Ile
30	Gln 65	Arg	Phe	Ile	Ser	Ala 70	Ser	Asn	His	Val	Glu 75	Phe	Thr	Lys	Cys	Val 80
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40	Asp	Lys	Asn 115	Leu	Leu	Ala	Tyr	Leu 120	Asp	Ile	Leu	Gln	His 125	Leu	Ser	Ser
	Tyr	Met 130	Lys	Arg	Thr	Ile	Phe 135	His	Ser	Leu	Leu	Leu 140	Tyr	Tyr	Ala	Ser
4 5	145					Trp 150					155					160
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	Ser	Asp	Asn	Gly 180	Gly	Ser	Asn	Asn	Ser 185	Asp	Lys	Thr	Ser	Ile 190	Ser	Gln
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		Leu 210	Leu	Thr	Asn	Val	Asn 215	Asn	Asp	His	His	Tyr 220	His	Leu	His	His
60	Ser 225	Ser	Ser	Ser	Ser	Lys 230	Thr	Thr	Asn	Thr	Asn 235	Ser	Pro	Asn	Ser	lle 240

	Ser	Lys	Thr	Ser	Ile 245		Gln	Ser	Ser	Val 250	Asn	Ala	Ser	Gly	Asn 255	Val
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		Thr			325					330					335	
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25		Arg	355					360					365			
25		Thr 370					375					380				
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		Ser			405	•				410				_	415	
35		Asp		420					425					430		
40			435					440					445			_
40		Ser 450					455					460				
45	465	Trp	GIÀ	ser	Ala	470	rys	Asn	Pro	Ser	475	Arg	His	Leu	Thr	His 480
		Leu			485					490					495	
50		Leu		500					505					510		
FF		Val	515					520					525			
5 5	Met	530					5 3 ₅					540				
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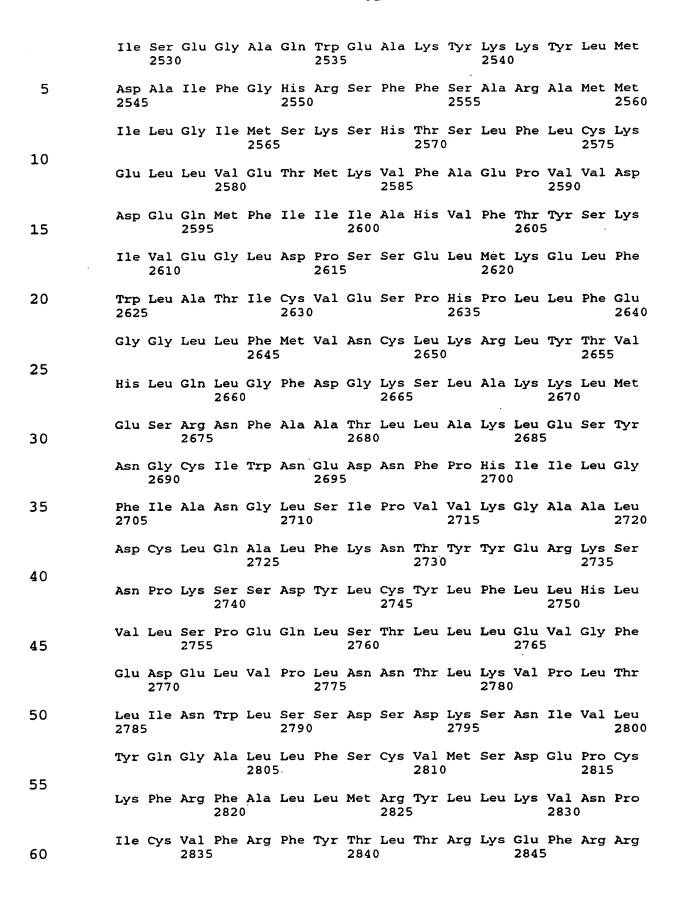
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	Ile	Leu 930	Asp	Ser	Asn	Gln	Arg 935	Leu	Gln	Val	Thr	Ala 940	Arg	Ala	Phe	Ile
10	Glu 945	Ile	Pro	Leu	Ser	Tyr 950	Ile	Ala	Thr	Phe	Glu 955	Asp	Ile	Asp	Asn	Asp 960
15	Leu	Asp	Pro	Arg	Val 965	Leu	Asn	Asp	His	Tyr 970	Leu	Leu	Cys	Thr	Tyr 975	Ala
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	Ser	Tyr 1010		Ser	Asn	Leu	Ala 1015		Lys	His	Asn	Leu 1020		Gln	Ala	Ile
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	Ile 1665		Ser	Ser	Ile	Asp 1670		Phe	Pro	Ile	Glu 1675		Val	Asp	Ile	Cys 1680
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•			1715	5		Glu		1720)				1725	5		
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					1765					1770)				1775	;
45				1780)	Asn			1785	;				1790)	
			1795	i		Asp		1800)				1805	5	_	
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					1845					1850					1855	
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	Ile Ile Val V 1905	al Thr Phe Thr 1910	Asn Cys Glu Tyr 191	Asn Asn Phe Val Met 5 1920
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	1955		1960	Ser Leu Ile Pro Glu 1965
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25		2005	2010	Ser Asn Thr Asp Gln 2015
30	20	20	2025	Ser Leu Glu Val Leu 2030
	Lys Asp Val Ai 2035	g Val Thr Leu	His Asp Ile Thr 2040	Leu Tyr Asp Lys Glu 2045
35	Lys Lys Lys Pr 2050	e Cys Pro Val 205		Gly Asn Lys Tyr Phe 2060
40	Gln Val Leu Hi 2065	s Glu Ile Pro 2070	Gln Leu Tyr Lys 207	Val Thr Val Ser Asn 2080
	Arg Thr Phe Se	r Ile Lys Phe 2085	Asn Asn Val Tyr 2090	Lys Ile Ser Asn Leu 2095
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	2115		2120	Cys Ser Pro Lys Tyr 2125
50	2130	213	5	Leu Lys Met Glu Glu 2140
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		2165	2170	Glu Val Gly Glu Ile 2175
60	Ile Ser His Le 21	u Ser Leu Val 80	Ile Leu Val Gly 2185	Leu Phe Asn Glu Asp 2190

	Asp	Leu	Val 219		Asn	Ile	Ser	Tyr 220		Leu	Leu	Val	Ala 220		Gln	Glu
5	Ala	Phe 221		Leu	Asp	Phe	Gly 221		Arg	Leu	His	Lys 222		Pro	Glu	Thr
	Tyr 222		Pro	Asp	Asp	Thr 223		Thr	Phe	Leu	Ala 223		Ile	Phe	Lys	Ala 2240
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15	Leu	Asp	Gly	Leu 226		Asn	Asp	Val	Ile 226	Pro 5	Gln	Glu	His	Ile 227		Thr
	Val	Val	Cys 227		Leu	Ser	Tyr	Trp 228		Pro	Asn	Leu	Tyr 2289		His	Val
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	Tyr 2309		Leu	Ile	Arg	Leu 2310		Val	Lys	Glu	Pro 2315		Phe	Thr	Thr	Ala 2320
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30	Asn	Val	Ile	Val 2340		Glu	Ile	Val	Ser 2349	His	Ala	Leu	Asp	Arg 2350	_	Ser
			2359	5				2360)	Ser			2365	5		
35		2370)				2375	5		Glu		2380)			
10	2385	5				2390)			Glu	2395	i				2400
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45				2420)				2425					2430)	
			2435	5				2440)	Glu			2445	;		_
50		2450)				2455	5		Leu		2460	1			
==	2465	;				2470	t			Ile	2475					2480
55					2485	•				Phe 2490	1			_	2495	5
60				2500					2505					2510)	
	ren	ASP	Leu 2515		inr	тÀ2	ASN	2520		Leu	ьeu	Met	Glu 2525		Gly	Ser



	Leu	Ser 2850		Leu	Glu	Gln	Ser 285		Glu	Ala	Val	Ala 2860		Ser	Phe	Glu
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	Phe	Asn	Asp		Met 2885		Glu	Leu	Leu	Lys 2890		Arg	Gly	Leu	Ser 2895	
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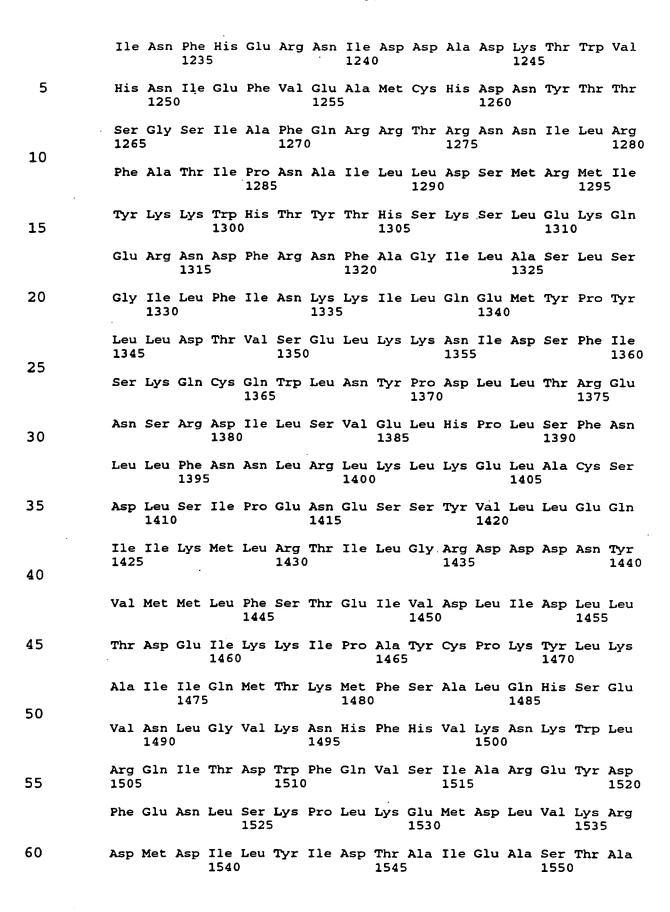
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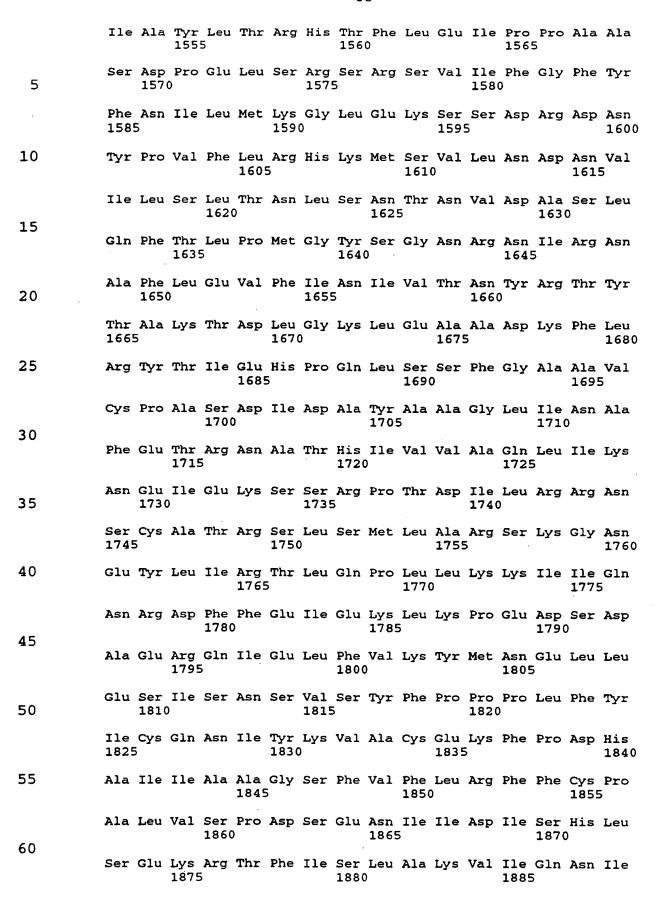
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	(vi)	ORI (A				: Sacc	haro	myce:	s ce:	revi	siae					
15	(xi)	SEQ	UENC:	E DE	SCRI	PTIO	N: S	EQ I	D NO	:4:						
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	Glu	Val 50	Glu	Glu	Tyr	Ser	Ser 55	Phe	Ile	Ser	Cys	Arg 60	Ser	Val	Leu	Ile
30	Asn 65	Val	Thr	Val	Ser	Arg 70	Asp	Ala	Asn	Ala	Met 75	Val	Glu	Gly	Thr	Let 80
35	Glu	Leu	Ile	Glu	Ser 85	Leu	Leu	Gln	Gly	His 90	Glu	Ile	Ile	Ser	Asp 95	Lys
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40	Ser	Asp	Ala 115	Leu	Glu	Tyr	Asn	Trp 120	Gln	Asn	Gln	Glu	Ser 125	Leu	His	Туз
	Asn	Asp 130	Ile	Ser	Thr	His	Val 135	Glu	His	Asp	Gln	Glu 140	Gln	Lys	Tyr	Arg
45	Pro 145	Lys	Leu	Asn	Ser	Ile 150	Leu	Pro	Asp	Tyr	Ser 155	Ser	Thr	His	Ser	Asr 160
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	Asn	Ile 210	Leu	Thr	Thr	Leu	Ser 215	Ser	Ser	Ile	Leu	Pro 220	Arg	His	Lys	Ser
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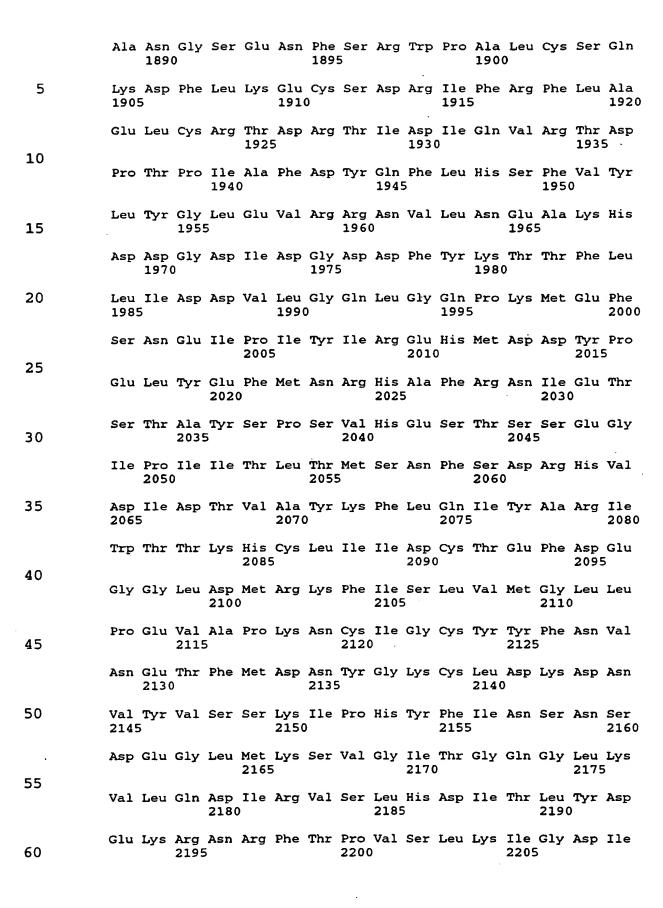
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10	Asn	Ser 290	Ile	Val	Pro	His	Phe 295	Asp	Leu	Phe	Ser	Phe 300	Ile	Tyr	Leu	Ser
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	Thr	Leu	Arg	Lys	Thr 325	Ile	Tyr	His	Cys	Leu 330	Leu	Leu	His	Tyr	Ser 335	Ala
20	Lys	Ala	Ile	Met 340	Phe	Trp	Ile	Met	Ala 345	Arg	Pro	Ala	Glu	Tyr 350	Tyr	Glu
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25		370					375					380				Asn
30	385				Ile	390					395					400
					Ser 405		•			410					415	
35				420					425					430		Leu
40			435		Ser			440					445	_		
40		450					455					460				Ser
45	465				Thr	470					475					480
					Ser 485					490					495	
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J J		530			Ser		535					540				
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	wsb	GIU	UIS	FIIG	Leu 565	SAT	vaı	IIIL	wrg	570	wsb	ASII	AGT	Leu	575	ьeu

	Tyr	Thr	His	Phe 580	Asp	Asp	Thr	Glu	Val 585	Leu	Pro	His	Thr	Ser 590	Val	Leu
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10	Leu	Asn 610	Ala	Thr	Ser	Phe	Lys 615	Tyr	Ile	Pro	Asp	Cys 620	Thr	Met	His	Arg
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25	Leu	Leu 690	Asp	Thr	Met	Arg	Ala 695	Leu	Leu	Ser	Phe	Phe 700	Thr	Met	Thr	Ser
	Ala 705	Val	Phe	Leu	Val	Asp 710	Arg	Asn	Leu	Pro	Ser 715	Val	Leu	Phe	Ala	Lys 720
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	Pro Ly		Leu Se	er Ser	Asp 935	Pro	Tyr	Leu	Ser	His 940	Leu	Val	Ala	Pro
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13	Val Gl	_	Pro G: 980	ln Thr	Glu	Ser	11e 985	Ser	Ala	Thr	Pro	Met 990	Ala	Ile
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25	Phe Me	t Lys	Ser T	nr Asn 103		Tyr	Ile	Gln	Glu 1035		Leu	Ile	Pro	Lys 1040
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35	Asn Il	e Asn 1075		sn Leu	Gln	Trp 1080		Ser	Gln	Asp	Phe 1089		Asn	Ile
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45	Tyr Gl	y Glu		sp Glu 125	Asn	Ile	Ser	Ile 113		Gly	Tyr	His	Leu 113	
	Cys Se	r Tyr	Thr Va 1140	al Thr	Leu	Phe	Ala 114		Gly	Leu	Phe	Asp 1150		Lys
50	Ile As	n Asn 1155		ys Arg	Gln	Ile 116		Leu	Asp	Ile	Thr 116		Lys	Phe
	Met Ly 11		Arg S	er His	Leu 117	_	Gly	Ile	Ala	Glu 118		Ser	His	His
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	Tyr Phe Gln 2210	Val Leu His	Glu Thr Pro 2215	Arg Gln Tyr 2220	Lys Ile Arg Asp
5	Met Gly Thr 2225	Leu Phe Asp 223		Asn Asp Val 2235	Tyr Glu Ile Ser 2240
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10	Phe Thr Val	Thr Phe Gln 2260	Asp Glu Arg 2265		Phe Ser Ser Pro 2270
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20	Ser Asn Asn 2305	Lys Val Lys 2310		Glu Arg Thr 2315	Ile Leu Leu Cys 2320
	His Leu Leu	Leu Val Ser 2325		Leu Phe Asp 2330	Glu Ser Lys Lys 2335
25	Met Lys Asn	Ser Ser Tyr 2340	Asn Leu Ile 2345		Glu Ala Ser Phe 2350
30	Gly Leu Asn 235		His Phe His . 2360	Arg Ser Pro	Glu Val Tyr Val 2365
	Pro Glu Asp 2370	Thr Thr Thr	Phe Leu Gly 2375	Val Ile Gly 2380	Lys Ser Leu Ala
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40	Ala Leu Lys	Asn Asn Val 2405		Val Tyr Ile 2410	Pro His Thr Ile 2415
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	Arg Asp Trp	Lys Lys Thr 2500	Ile Ser Leu 1 2505	Leu Thr Val	Leu Pro Thr Thr 2510
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	Leu	Leu Va 3010	l Thr	His	Ser	Ġlu 3015	Ser	Asn	Ser	Leu	Glu 3020		Leu	His	Glu
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50	(i)	(B) T (C) S	CE CHA ENGTH: YPE: a TRANDE OPOLOG	870 mino DNES:	ami acio S: s:	no a d ingl	cids	•							
55	(ii)	MOLECU	LE TYP	E: p	rote	in									
	(vi)	ORIGIN (A) O	AL SOU RGANIS		omo:	sapi	ens								
60															

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		Lys	Thr 290	Lys	Asp	Ala	Phe	Tyr 295	Lys	Asn	Ile	Val	L ys 300	Lys	Gly	Tyr	Leu
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	Glu	Gly	Ser	Asp	Ala 325	Gln	Leu	Ile	Tyr	Phe 330	Glu	Ser	Glu	Lys	Arg 335	Ala
5	Thr	Lys	. Pro	Lys 340	Gly	Leu	Ile	Asp	Leu 345	Ser	Val	Cys	Ser	Val 350	Tyr	Val
	Val	His	Asp 355	Ser	Leu	Phe	Gly	Arg 360	Pro	Asn	Cys	Phe	Gln 365	Ile	Val	Val
10	Gln	His 370	Phe	Ser	Glu	Glu	His 375	Tyr	Ile	Phe	Tyr	Phe 380	Ala	Gly	Glu	Thr
15	Pro 385	Glu	Gln	Ala	Glu	Asp 390	Trp	Met	Lys	Gly	Leu 395	Gln	Ala	Phe	Cys	Asn 400
	Leu	Arg	Lys	Ser	Ser 405	Pro	Gly	Thr	Ser	Asn 410	Lys	Arg	Leu	Arg	Gln 415	Val
20	Ser	Ser	Leu	Val 420	Leu	His	Ile	Glu	Glu 425	Ala	His	Lys	Leu	Pro 430	Val	Lys
	His	Phe	Thr 435	Asn	Pro	Tyr	Cys	Asn 440	Ile	Tyr	Leu	Asn	Ser 445	Val	Gln	Val
25	Ala	Lys 450	Thr	His	Ala	Arg	Glu 455	Gly	Gln	Asn	Pro	Val 460	Trp	Ser	Glu	Glu
30	Phe 465	Val	Phe	Asp	Asp	Leu 470	Pro	Pro	Asp	Ile	Asn 475	Arg	Phe	Glu	Ile	Thr 480
					485					Asp 490					495	
35				500					505	Gly				510		
40			515					520		Lys			525		_	
40		530					5 35			Glu		540				
45	545					550				Leu	555					560
					565					Gln 570					57 5	
50	·			580					585	Glu				590		
5 5			595					600		Ser			605			
55		610					615			Ser		620				
60	625					630				His	635					640
	TTE	ьeu	ъÀг	TTE	Met 645	GIU	ser	nys	GIN	Ser 650	Cys	Glu	Leu	Ser	Pro 655	Ser

	Lys	Leu	Glu		Asn	Glu	Ásp	Val		Thr	Asn	Leu	Thr		Leu	Leu
5	Asn	Ile	Leu	660 Ser	Glu	Leu	Val	Glu	665	Ile	Phe	Met	Δla	670 Ser	Glu	Tle
J			675			202	•41	680	2,0		1110		685	Der	014	110
10	Leu	Pro 690	Pro	Thr	Leu	Arg	Tyr 695	Ile	Tyr	Gly	Cys	Leu 700	Gln	Lys	Ser	Val
10	Gln 705	His	Lys	Trp	Pro	Thr 710	Asn	Thr	Thr	Met	Arg 715	Thr	Arg	Val	Val	Ser 720
15	Gly	Phe	Val	Phe	Leu 725	Arg	Leu	Ile	Cys	Pro 730	Ala	Ile	Leu	Asn	Pro 735	Arg
	Met	Phe	Asn	11e 740	Ile	Ser	Asp	Ser	Pro 745	Ser	Pro	Ile	Ala	Ala 7 50	Arg	Thr
20	Leu	Ile	Leu 755	Val	Ala	Lys	Ser	Val 760	Gln	Asn	Leu	Ala	Asn 765	Leu	Val	Glu
25	Phe	Gly 770	Ala	Lys	Glu	Pro	Tyr 775	Met	Glu	Gly	Val	Asn 780	Pro	Phe	Ile	Lys
23	Ser 785	Asn	Lys	His	Arg	Met 790	Ile	Met	Phe	Leu	Asp 795	Glu	Leu	Gly	Asn	Val 800
30	Pro	Glu	Leu	Pro	Asp 805	Thr	Thr	Glu	His	Ser 810	Arg	Thr	Asp	Leu	Ser 815	Arg
	Asp	Leu	Ala	Ala 820	Leu	His	Glu	Ile	Cys 825	Val	Ala	His	Ser	Asp 830	Glu	Leu
35	Arg	Thr	Leu 835	Ser	Asn	Glu	Arg	Gly 840	Ala	Gln	Gln	His	Val 845	Leu	Lys	Ļys
40	Leu	Leu 850	Ala	Ile	Thr	Glu	Leu 855	Leu	Gln	Gln	Lys	Gln 860	Asn	Gln	Tyr	Thr
40	L ys 865	Thr	Asn	Asp _.	Val	A rg 8 70										
45	(2) INFOR	ITAM	ON F	OR S	EQ I	D NC	:6:									
	(i)	(A) (B)	LEN TYP	GTH: E: a	766 mino	ami aci	ď	cids	;							
50			STR				ingl r	.e		٠						
	(ii)	MOLE	CULE	TYP	E: p	rote	in									
55	(vi)						osac	char	omyc	es p	ombe					
	(xi)	SEQU	ENCE	DES	CRIP	TION	: SE	Q ID	NO:	6 :						
60	Met 1	Thr	Lys		His 5	Ser	Gly	Thr	Leu	Ser 10	Ser	Ser	Val	Leu	Pro 15	Gln

	Thr	Asn	Arg	Leu 20	Ser	Leu	Leu	Arg	Asn 25	Arg	Glu	Ser	Thr	Ser 30	Val	Leu
5	Tyr	Thr	Ile 35	Asp	Leu	Asp	Met	Glu 40	Ser	Asp	Val	Glu	Asp 45	Ala	Phe	Phe
	His	Leu 50	Asp	Arg	Glu	Leu	His 55	Asp	Leu	Lys	Gln	Gln 60	Ile	Ser	Ser	Gln
10	Ser 65	Lys	Gln	Asn	Phe	Val 70	Leu	Glu	Arg	Asp	Val 75	Arg	Tyr	Leu	Asp	Ser 80
15	Lys	Ile	Ala	Leu	Leu 85	Ile	Gln	Asn	Arg	Met 90	Ala	Gln	Glu	Glu	Gln 95	His
13	Glu	Phe	Ala	Lys 100	Arg	Leu	Asn	Asp	Asn 105	Tyr	Asn	Ala	Val	Lys 110	Gly	Ser
20	Phe	Pro	Asp 115	Asp	Arg	Lys	Leu	Gln 120	Leu	Tyr	Gly	Ala	Leu 125	Phe	Phe	Leu
	Leu	Gln 130	Ser	Glu	Pro	Ala	Tyr 135	Ile	Ala	Ser	Leu	Val 140	Arg	Arg	Val	Lys
25	Leu 145	Phe	Asn	Met	Asp	Ala 150	Leu	Leu	Gln	Ile	Val 155	Met	Phe	Asn	Ile	Tyr 160
30	Gly	Asn	Gln	Tyr	Glu 165	Ser	Arg	Glu	Glu	His 170	Leu	Leu	Leu	Ser	Leu 175	Phe
30	Gln	Met	Val	Leu 180	Thr	Thr	Glu	Phe	Glu 185	Ala	Thr	Ser	Asp	Val 190	Leu	Ser
35	Leu	Leu	Arg 195	Ala	Asn	Thr	Pro	Val 200	Ser	Arg	Met	Leu	Thr 205	Thr	Tyr	Thr
	Arg	Arg 210	Gly	Pro	Gly	Gln	Ala 215	Tyr	Leu	Arg	Ser	Ile 220	Leu	Tyr	Gln	Cys
40	225					230					235		_	Ile		240
45	Leu	Ser	Val	Tyr	Arg 245	Tyr	Leu	Val	Asn	Thr 250	Gly	Gln	Leu	Ser	Pro 255	Ser
	Glu	Asp	Asp	Asn 260	Leu	Leu	Thr	Asn	Glu 265	Glu	Val	Ser	Glu	Phe 270	Pro	Ala
50	Val	Lys	Asn 275	Ala	Ile	Gln	Glu	Arg 280	Ser	Ala	Gln	Leu	Leu 285	Leu	Leu	Thr
	Lys	Arg 290	Phe	Leu	Asp	Ala	Val 295	Leu	Asn	Ser	Ile	Asp 300	Glu	Ile	Pro	Tyr
55	Gly 305	Ile	Arg	Trp	Val	Cys 310	Lys	Leu	Ile	Arg	Asn 315	Leu	Thr	Asn	Arg	Leu 320
60	Phe	Pro	Ser	Ile	Ser 325	Asp	Ser	Thr	Ile	Cys 330	Ser	Leu	Ile	Gly	Gly 335	Phe
-	Phe	Phe	Leu	Arg 340	Phe	Val	Asn	Pro	Ala 345	Ile	Ile	Ser	Pro	Gln 350	Thr	Ser

5	Met	Leu	Leu 355	Asp	Ser	Cys	Pro	Ser 360	Asp	Asn	Val	Arg	L ys 365	Thr	Leu	Ala
3	Thr	Ile 370	Ala	Lys	Ile	Ile	Gln 375	Ser	Val	Ala	Asn	Gly 380	Thr	Ser	Ser	Thr
10	Lys 385	Thr	His	Leu	Asp	Val 390	Ser	Phe	Gln	Pro	Met 395	Leu	Lys	Glu	Tyr	Glu 400
	Glu	Lys	Val	His	Asn 405	Leu	Leu	Arg	Lys	Leu 410	Gly	Asn	Val	Gly	Asp 415	Phe
15	Phe	Glų	Ala	Leu 420	Glu	Leu	Asp	Gln	Tyr 425	Ile	Ala	Leu	Ser	Lys 430	Lys	Ser
20	Leu	Ala	Leu 435	Glu	Met	Thr	Val	Asn 440	Glu	Ile	Tyr	Leu	Thr 445	His	Glu	Ile
	Ile	Leu 450	Glu	Asn	Leu	Asp	Asn 455	Leu	Tyr	Asp	Pro	Asp 460	Ser	His	Val	His
25	Leu 465	Ile	Leu	Gln	Glu	Leu 470	Gly	Glu	Pro	Cys	Lys 475	Ser	Val	Pro	Gln	Glu 480
	Asp	Asn	Cys	Leu	Val 485	Thr	Leu	Pro	Leu	Tyr 490	Asn	Arg	Trp	Asp	Ser 495	Ser
30			_	500					505				Glu	510		
35	Tyr	Val	Asp 515	Ala	Lys	Thr	Leu	Phe 520	Ile	Gln	Leu	Leu	Arg 525	Leu	Leu	Pro
	Ser	Gly 530	His	Pro	Ala	Thr	Arg 535	Val	Pro	Leu	Asp	Leu 540	Pro	Leu	Ile	Ala
40	Asp 545	Ser	Val	Ser	Ser	Leu 550	Lys	Ser	Met	Ser	Leu 555	Met	Lys	Lys	Gly	Ile 560
	_				565					570			Arg	•	57 5	
45	Lys	Glu	Asn	Arg 580	Tyr	Glu	Pro	Leu	Thr 585	Ser	Glu	Val	Glu	Lys 590	Glu	Phe
50	Ile	Asp	Leu 595	Asp	Ala	Leu	Tyr	Glu 600	Arg	Ile	Arg	Ala	Glu 605	Arg	Asp	Ala
	Leu	Gln 610	Asp	Val	His	Arg	Ala 615	Ile	Cys	Asp	His	Asn 620	Glu	Tyr	Leu	Gln
5 5	Thr 625	Gln	Leu	Gln	Ile	Tyr 630	Gly	Ser	Tyr	Leu	Asn 635	Asn	Ala	Arg	Ser	Gln 640
	Ile	Lys	Pro	Ser	His 645	Ser	Asp	Ser	Lys	Gly 650	Phe	Ser	Arg	Gly	Val 655	Gly
60	Val	Val	Gly	Ile 660	Lys	Pro	Lys	Asn	11e 665	Lys	Ser	Ser	Asn	Thr 670	Val	Lys

	Leu Se	r Ser 675		Gln	Leu	Lys	Lys 680	Glu	Ser	Val	Leu	Leu 685	Asn	Cys	Thr
5	Ile Pr 69		Phe	Asn	Val	Ser 695	Asn	Thr	Tyr	Phe	Thr 700	Phe	Ser	Ser	Pro
	Ser Th	r Asp	Asn	Phe	Val 710	Ile	Ala	Val	Tyr	Gln 715	Arg	Gly	His	Ser	Lys 720
10	Val Le	u Val	Glu	Val 725	Cys	Ile	Cys	Leu	Asp 730	Asp	Val	Leu	Gln	Arg 735	Arg
15	Tyr Al	a Ser	Asn 740	Pro	Val	Val	Asp	Leu 745	Gly	Phe	Leu	Thr	Phe 750	Glu	Ala
	Asn Ly	5 Leu 755	Tyr	His	Leu	Phe	Glu 760	Gln	Leu	Phe	Leu	Arg 765	Lys		

WHAT IS CLAIMED IS:

- A method of blocking a Ras-induced effect on a
 cell, comprising a step of introducing a GTPase Activating (GAP) protein to said cell.
 - 2. A method of Claim 1, wherein said Ras is an oncogenic Ras.

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- 3. A method of Claim 1, wherein said Ras substantially lacks GTPase activity.
- 4. A method of Claim 1, wherein said effect is induction of cell proliferation or transformation.
 - 5. A method of treating an oncogenic Ras transformed cell comprising the step of introducing to said cell a GAP protein capable of suppressing the
- 20 transformation of said cell.
 - 6. The method of either Claim 1 or 5, wherein said cell is a eukaryotic cell, including a mammalian cell, including a human cell.

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- 7. The method of either Claim 1 or 5, wherein said step of introducing is by expression of a nucleic acid encoding said GAP protein.
- 30 8. A method for the manufacture of a pharmaceutical composition for treating an oncogenic Ras transformed cell comprising admixing a GAP protein capable of suppressing the transformation of said cell with a pharmaceutically acceptable carrier.

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9. The method of any of Claims 1,5or 8 wherein said GAP protein binds to said Ras protein with a Kd of less than 200 nM.

- 10. The method of any of Claims 1,5 or 8 wherein said GAP protein is selected from the group of:
 - a) a fragment of a mammalian GAP protein;
 - b) a fragment of a mammalian NF1-GRD protein;
 - c) a homologue or mimetic of a or b; and
 - d) the proteins defined by SEQ ID NO: 1 or SEQ ID NO: 2.
- 10 11. The method of any of Claims 1, 5 or 8 wherein said GAP protein is selected from the group of:
 - a) a fragment of a mammalian GAP protein having a wild type sequence, including a human GAP protein; and
- b) a fragment of a mutant mammalian GAP protein having a sequence with an amino acid substitution at a position corresponding to a position 1063 through 1651 of NF1 or the corresponding region of other GAP proteins.

- 12. A method of Claim 11, wherein said substitution is a conservative substitution.
- 13. The method of any of Claims 1, 5 or 8 wherein said GAP protein interacts with Ras and blocks interaction of an effector molecule which binds to Ras at a position from 32 to 40 or from 59 to 65.
- 14. A method of Claim 2, wherein said GAP protein30 does not block signal transduction of non-oncogenic Ras.
- 15. A method of either of Claim 1 or 2, further comprising the steps of identifying the responsible oncogenic Ras and selecting said GAP protein which blocks said identified oncogenic Ras.

- 16. A method of identifying appropriate GAP proteins useful for treating a mutated Ras-induced condition of a eukaryote cell comprising the steps of:
 - a) identifying the mutated Ras which induces said condition; and
 - b) screening various GAP variants for specific variants which are capable of blocking said condition.
- 10 17. A method of Claim 16, wherein said eukaryote cell is a mammalian cell, including a human cell.
- 18. A method of 16, further comprising additional screening to determine which GAP variants have minimal effect on non-mutated Ras effects.
 - 19. A GAP protein capable of blocking transformation of a cell, where said transformation is due to an oncogenic Ras.

- 20. A protein of Claim 19, wherein said GAP is selected from the group of:
 - a) a fragment of a mammalian GAP protein;
 - b) a fragment of a mammalian NF1-GRD protein;
- c) a homologue or mimetic of a or b; and
 - d) a protein defined by SEQ ID NO: 1 or SEQ ID NO:2.
- 21. A protein of Claim 19, selected from the group 30 of:
 - a fragment of a mammalian GAP protein having a wild type sequence, including a human GAP protein; and
- b) a fragment of a mutant mammalian GAP protein
 having a sequence with an amino acid
 substitution at a position corresponding to a
 position from 1063 through 1651 or the
 corresponding region of other GAP proteins.

- 22. A protein of Claim 21, wherein said substitution is a conservative substitution.
- 5 23. A protein of Claim 19, wherein said protein interacts with Ras and blocks interaction of an effector molecule which binds to Ras at a position from 32 to 40 or from 59 to 65.
- 10 24. A protein of Claim 19, wherein said cell is a eukaryotic cell, including a mammalian cell, including a human cell.
- 25. A protein of Claim 19, wherein said oncogenic 15 Ras substantially lacks GTPase activity.
 - 26. A protein of Claim 19, which binds to said Ras protein with a Kd of less than 200 nM.
- 20 27. A protein of Claim 19, wherein said protein interferes with interaction of Ras•GTP with an effector compound.
- 28. An isolated nucleic acid encoding a protein 25 normally expressed as a protein of Claim 19.
- 29. A pharmaceutical composition for treating an oncogenic Ras transformed cell comprising a GAP protein capable of suppressing the transformation of said cell and a pharmaceutically carrier.
 - 30. The pharmaceutical composition of claim 29 wherein the GAP protein binds to said Ras protein with a Kd of less than 200 nM.
 - 31. The pharmaceutical composition of claim 29 wherein said GAP protein is selected from the group of:

 a) a fragment of a mammalian GAP protein;

- b) a fragment of a mammalian NF1-GRD protein;
- c) a homologue or mimetic of a or b; and
- d) the proteins defined by SEQ ID NO: 1 or SEQ ID NO: 2.

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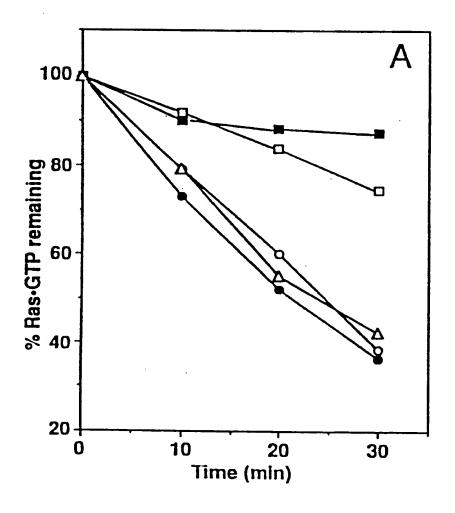
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- 32. The pharmaceutical composition of claim 29 wherein said GAP protein is selected from the group of:
 - a) a fragment of a mammalian GAP protein having a wild type sequence, including a human GAP protein; and
 - b) a fragment of a mutant mammalian GAP protein having a sequence with an amino acid substitution at a position corresponding to a position 1063 through 1651 of NF1 or the corresponding region of other GAP proteins.
- 33. The pharmaceutical composition of claim 29 wherein said GAP protein interacts with Ras and blocks 20 interaction of an effector molecule which binds to Ras at a position from 32 to 40 or from 59 to 65.
- 34. The use of a GAP protein capable of suppressing the transformation of an oncogenic Ras transformed cell and a pharmaceutically carrier for treating said oncogenic Ras transformed cell.
- 35. The use of a GAP protein capable of suppressing the transformation of an oncogenic Ras transformed cell for the manufacture of a medicament for treating said oncogenic Ras transformed cell.
- 36. The use of either Claim 34 or 35 in which the GAP protein binds to said Ras protein with a Kd of less than 200 nM.
 - 37. The use of either Claim 34 or 35 in which the GAP protein is selected from the group of:

- a) a fragment of a mammalian GAP protein;
- b) a fragment of a mammalian NF1-GRD protein;
- c) a homologue or mimetic of a or b; and
- d) the proteins defined by SEQ ID NO: 1 or SEQ ID NO: 2.
- 38. The use of either Claim 34 or 35 in which the GAP protein is selected from the group of:
- a) a fragment of a mammalian GAP protein having a wild type sequence, including a human GAP protein; and
 - b) a fragment of a mutant mammalian GAP protein having a sequence with an amino acid substitution at a position corresponding to a position 1063 through 1651 of NF1 or the corresponding region of other GAP proteins.
- 39. The use of either Claim 34 or 35 in which the
 20 GAP protein interacts with Ras and blocks interaction of an
 effector molecule which binds to Ras at a position from 32
 to 40 or from 59 to 65.

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FIGURE 1A



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FIGURE 1B

